

Long-term Patterns in Lake Champlain Plankton: the role of Invasive Species and Climate Change

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Jillian Zajac, Joe Connelly, Sarah Glancy

Undergrads- Carrienne Pershyn, Amanda Greene, Amanda Groves, Allison
Waring, Lindsay Paul, Gabriella Doud, Erin Moseman
and a whole host of other SUNY Plattsburgh students



Introduction



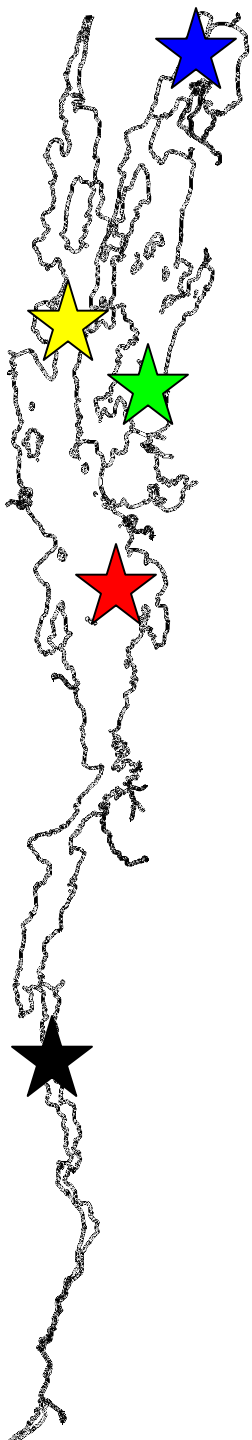
- 200 km in length,
- surface area 1130 km²
- mean depth 19.4 meters
- 3 year water residence time



Long-Term Water Quality and Biological Monitoring Project

Supported by the Lake Champlain Basin Program (U.S. EPA)
Collaborators- Fred Dunlap (NY DEC), Pete Stengel, Angela Shambaugh, Eric Smeltzer (VT DEC)





Zooplankton trends: NYDEC Monitoring program

5 sites

Summer (July/August) 1992-2009

Station #	Latitude (North)	Longitude (West)	Depth (meters)
 50	45 00.80	73 10.43	04
 36	44 45.37	73 21.30	50
 34	44 42.49	73 13.61	50
 19	44 28.26	73 17.95	100
 04	43 57.10	73 24.47	10

Rotifers

Wheel animals



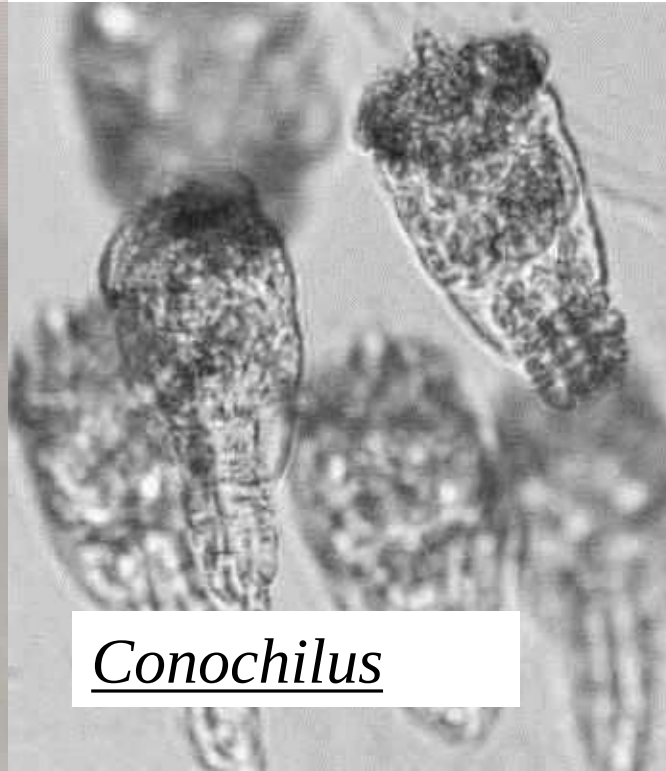
Asplanchna



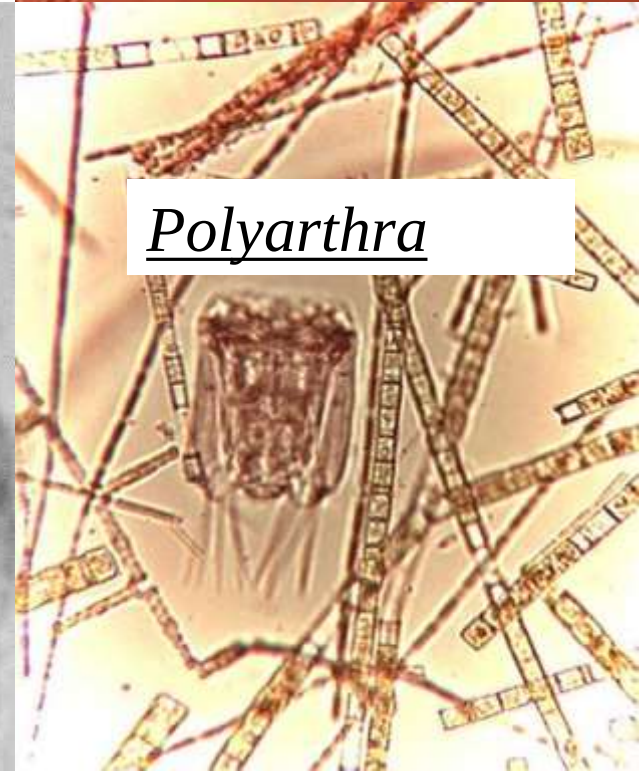
Kellicottia



Kerratella



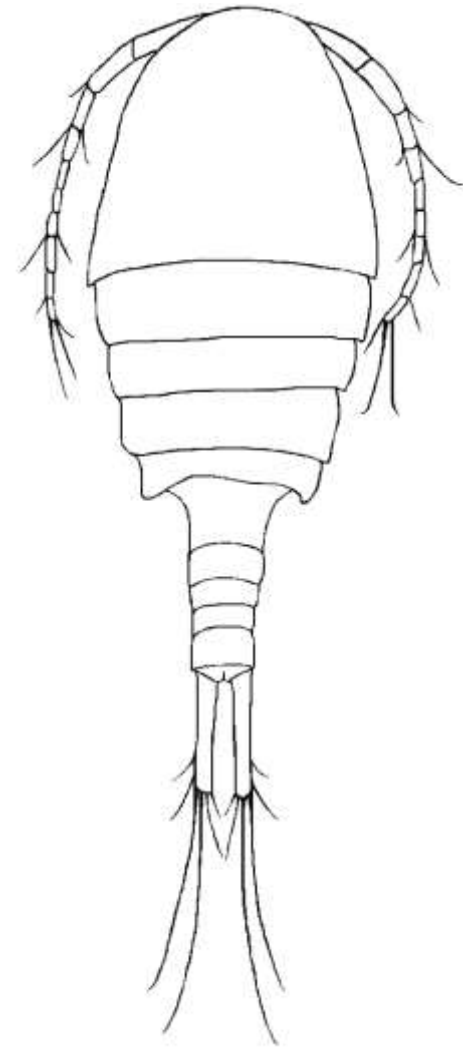
Conochilus



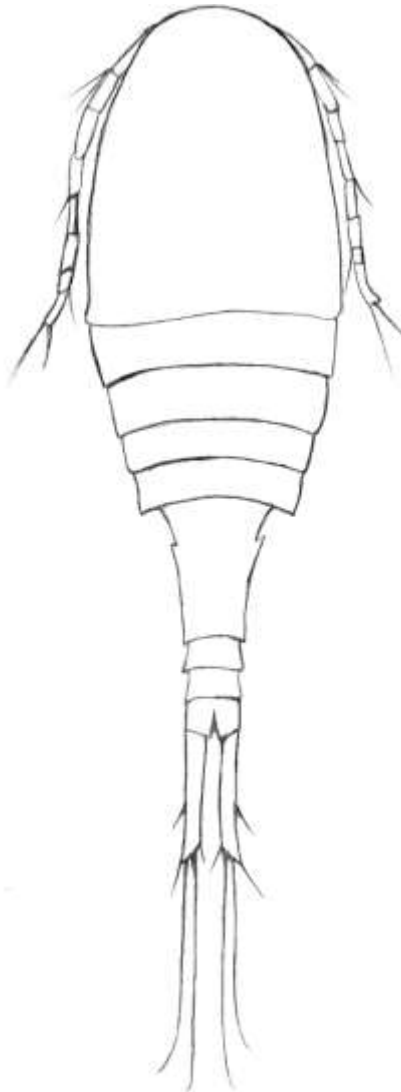
Polyarthra

Key to Zooplankton of Lake Champlain

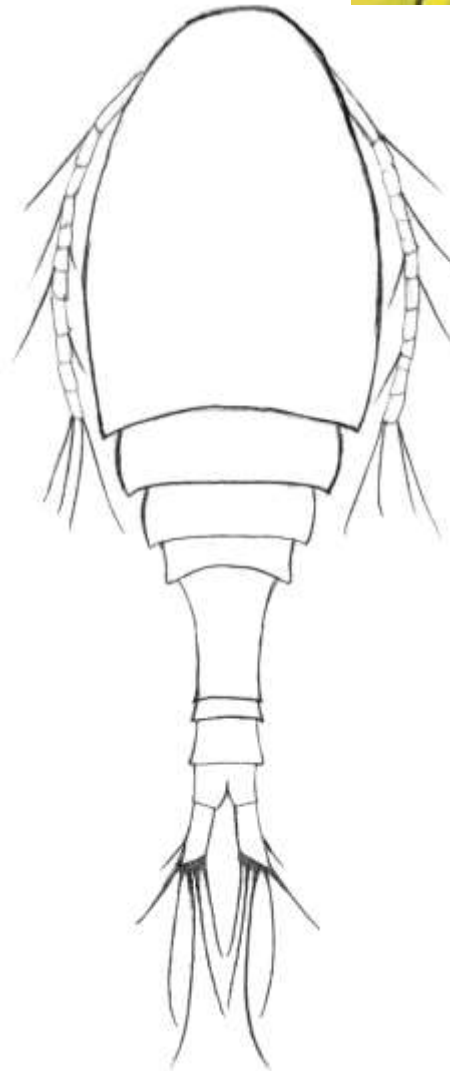
Copepods



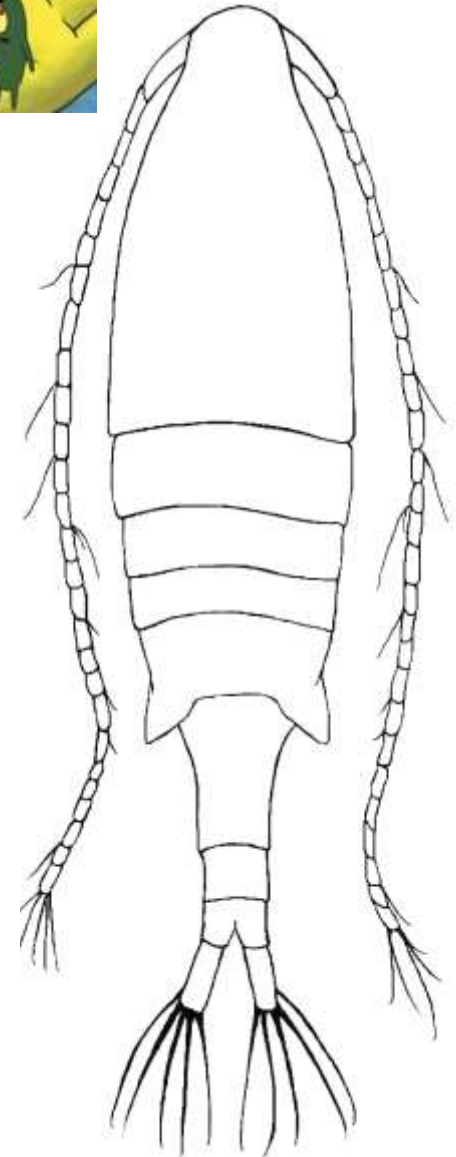
Acanthocyclops



Diacyclops

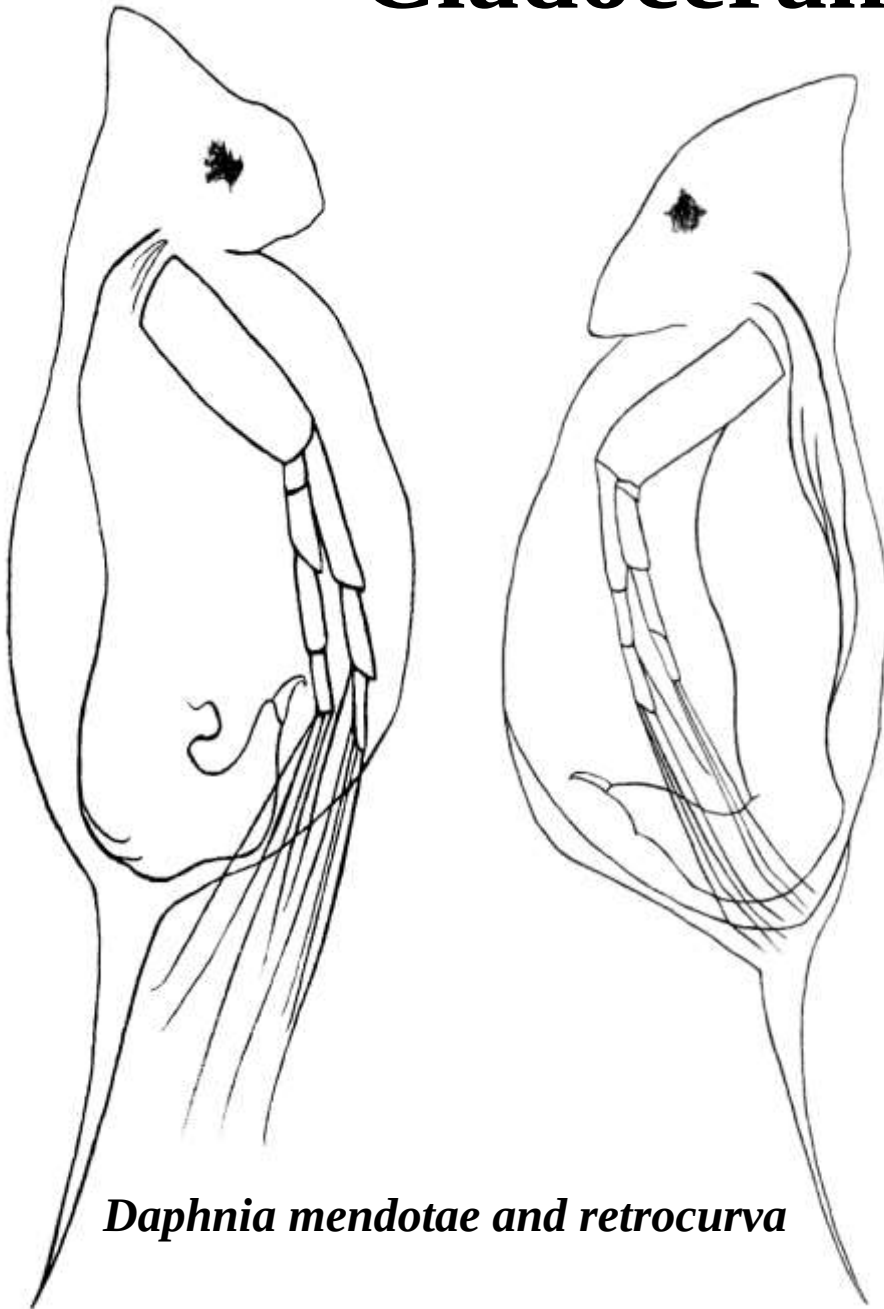


Mesocyclops



Leptodiaptomus

Cladocerans



Daphnia mendotae and retrocurva



Diaphanasoma

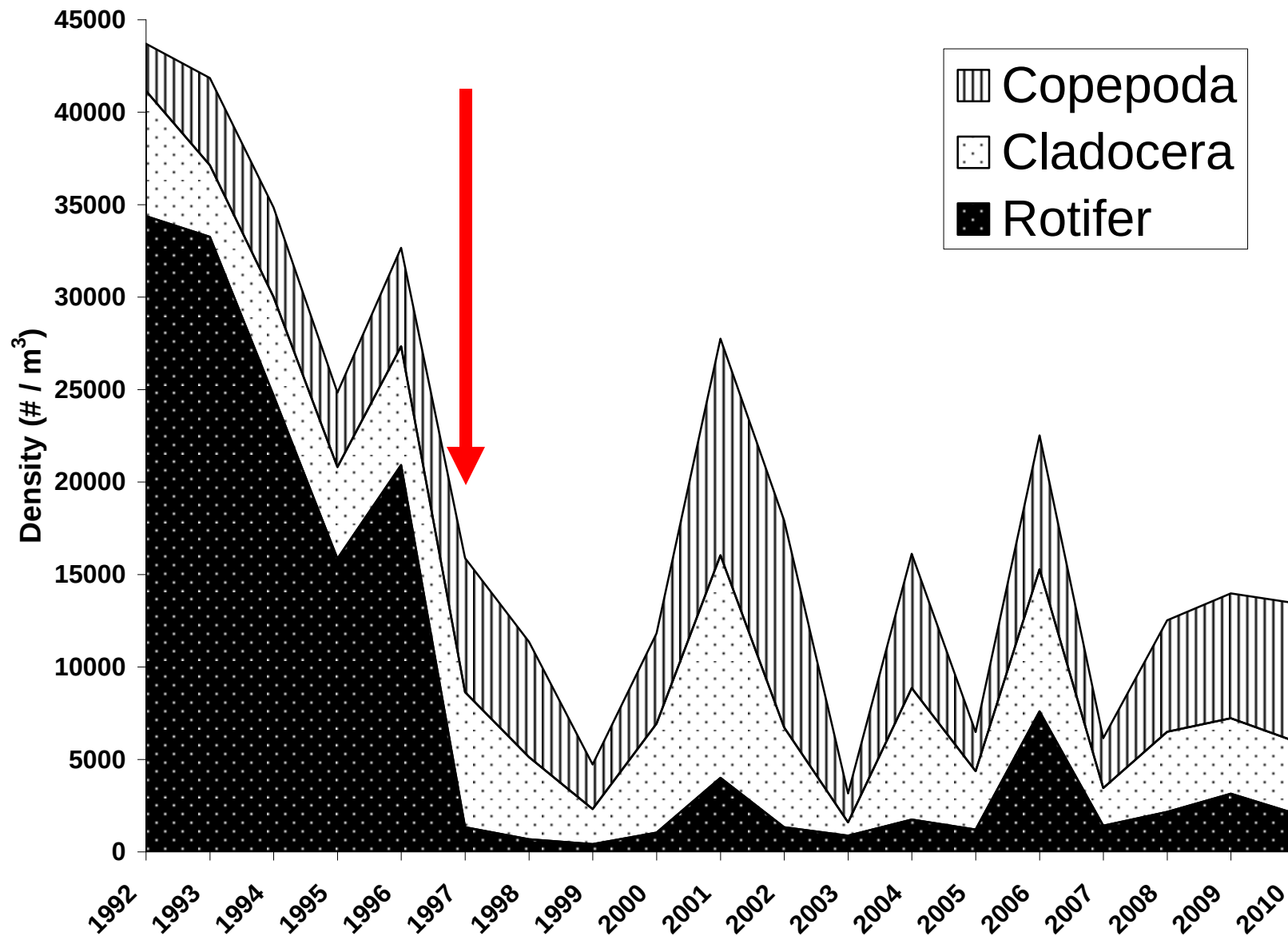


Eubosmina



Bosmina

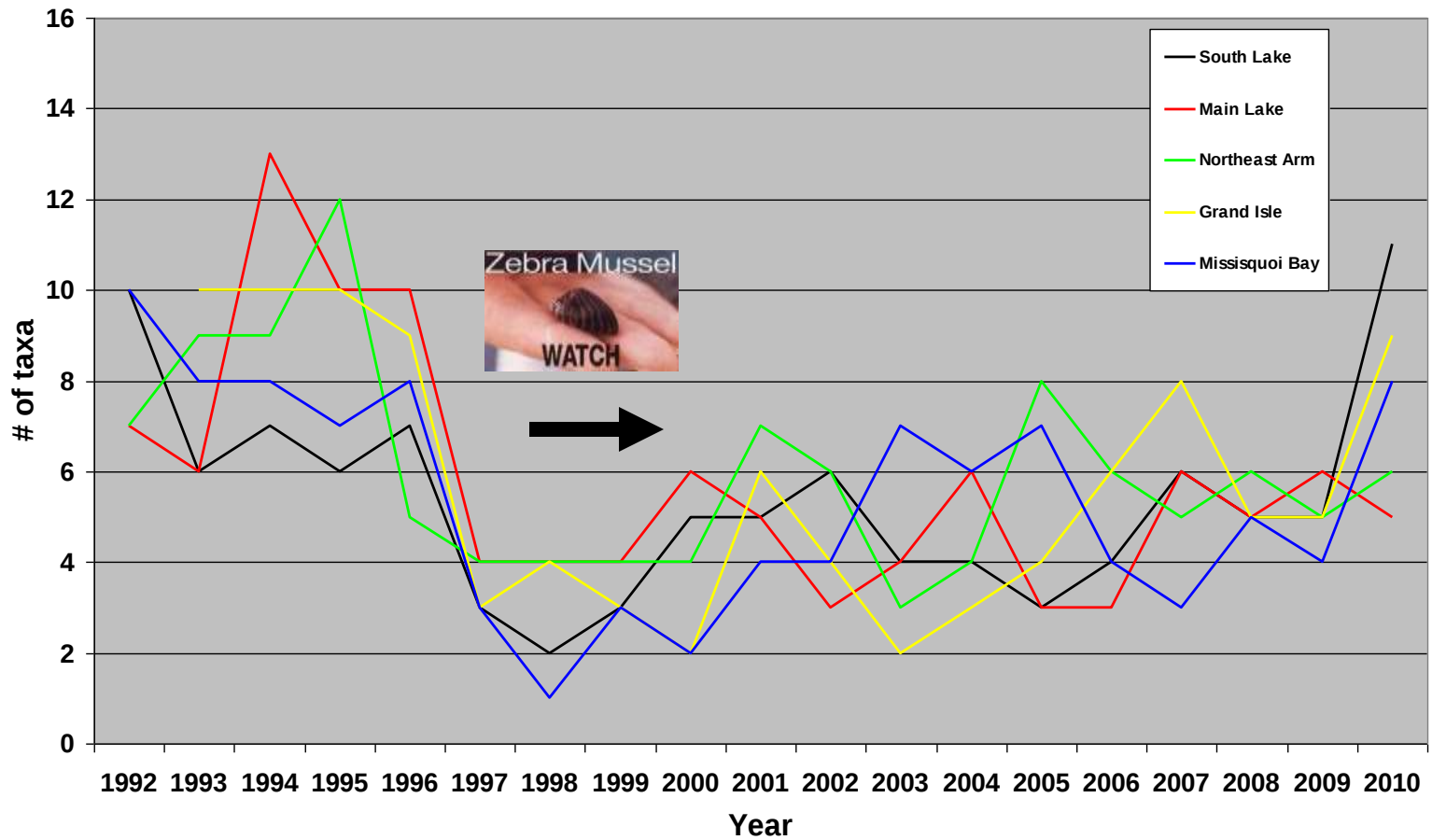
Deep lake study sites

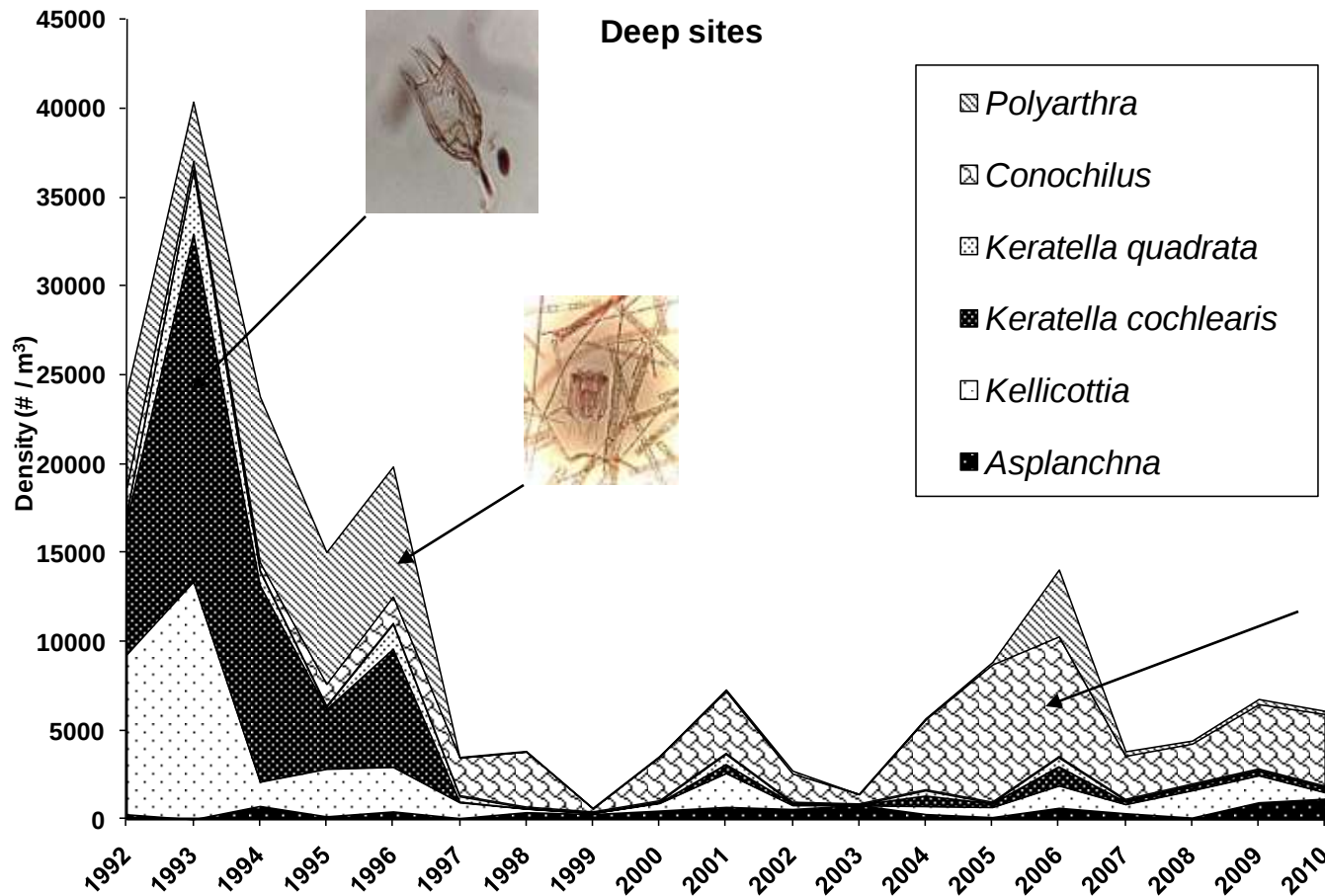
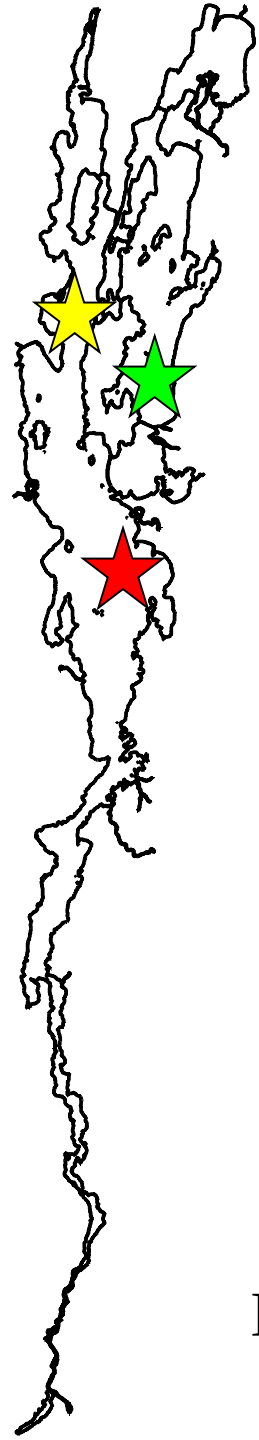


Mihuc et al. 2012



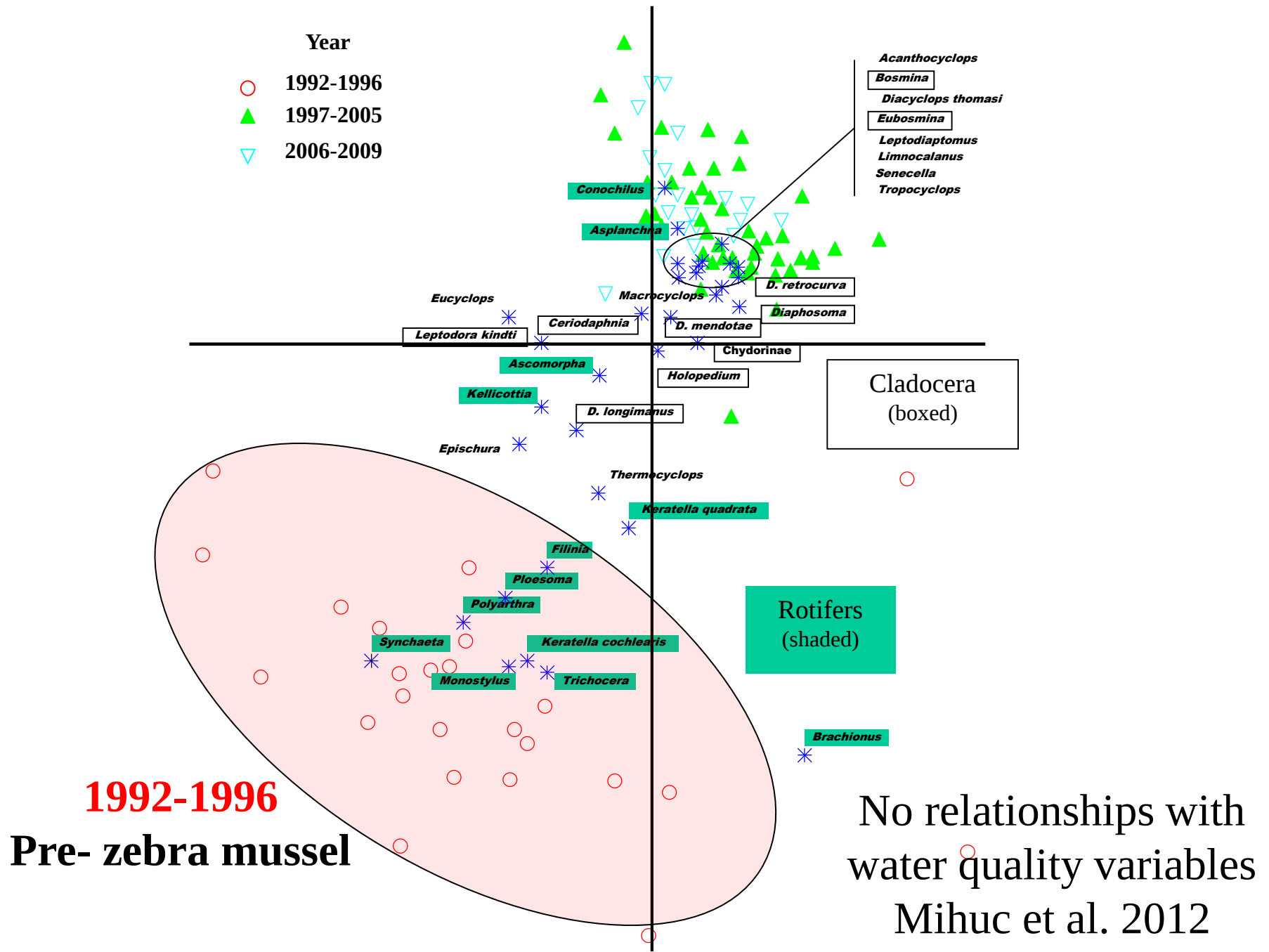
Rotifer taxa richness





Conochilus **SURVIVED**

Deep sites Lake Champlain
Mihuc et al. 2012





Long-term patterns in Lake Champlain's zooplankton: 1992–2010

Timothy B. Mihuc^{a,*}, Fred Dunlap^b, Casey Binggeli^a, Luke Myers^a, Carrienne Pershyn^a,
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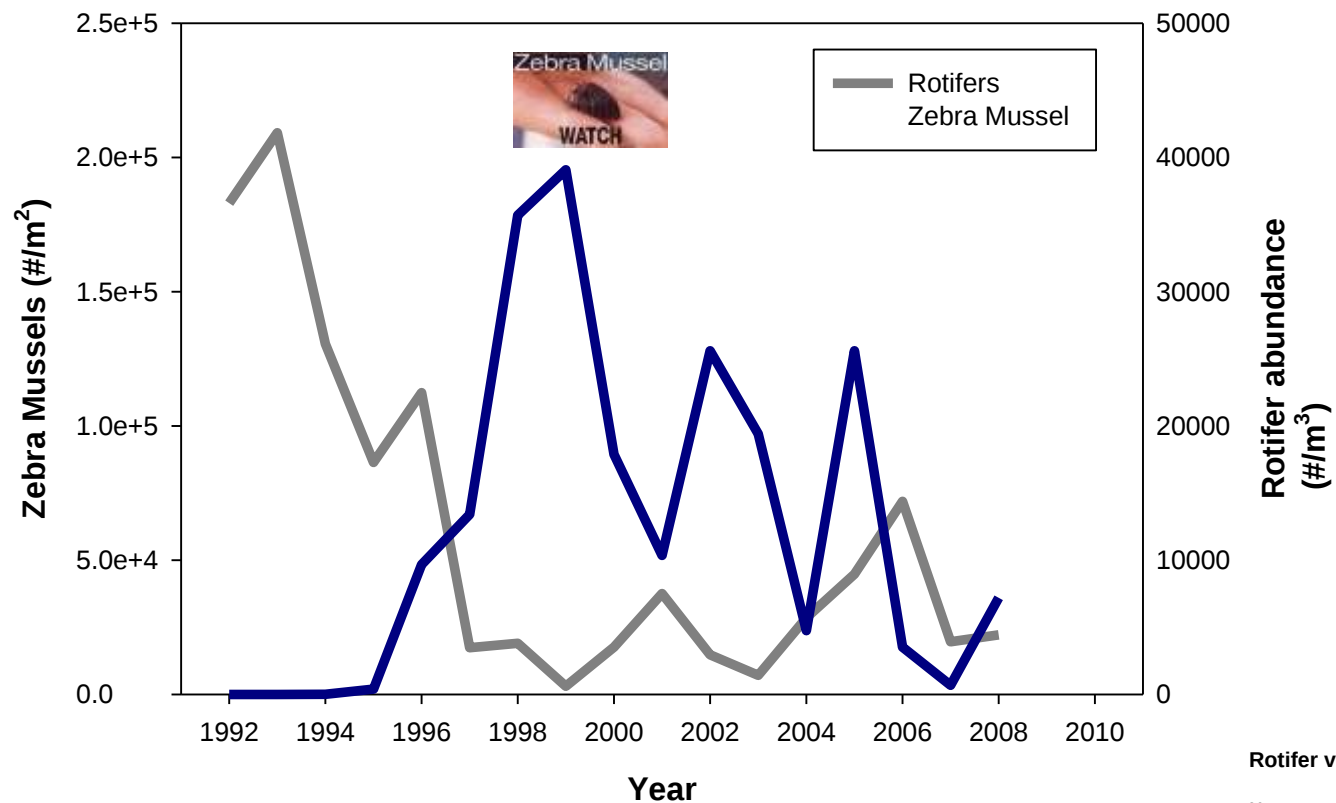
Invasive species

ABSTRACT

We examined patterns in Lake Champlain zooplankton abundance from 1992 to 2010 using summer data from five study sites. Rotifer abundance ($\#/m^3$) for many common taxa such as *Polyarthra*, *Kellicottia*, and *Keratella* declined lakewide in the mid-1990s which coincided with the invasion of zebra mussels (*Dreissena polymorpha*) into Lake Champlain. The only rotifer to increase in density following zebra mussel invasion was *Conochilus* which is a colonial species. Long-term shifts in copepod and cladoceran community composition can be attributed to the arrival of another invasive species in 2004–2005, the alewife (*Alosa pseudoharengus*). Our results support previous findings that alewife predation can impact larger bodied zooplankton within temperate lake systems. Following alewife invasion into Lake Champlain, body length of *Leptodiatomus* and *Daphnia retrocurva* decreased to a size at or below known alewife feeding preferences. In addition, smaller bodied copepods (primarily *Diacyclops thomasi*) have increased in abundance since alewife invasion while juvenile copepods have declined. Our results suggest that post-alewife zooplankton patterns are most likely due to alewife size-selective feeding strategies. Observed long-term changes in zooplankton community structure have potential implications for the lake's food web dynamics, particularly recent declines in large bodied zooplankton which may release smaller plankton from top-down control.

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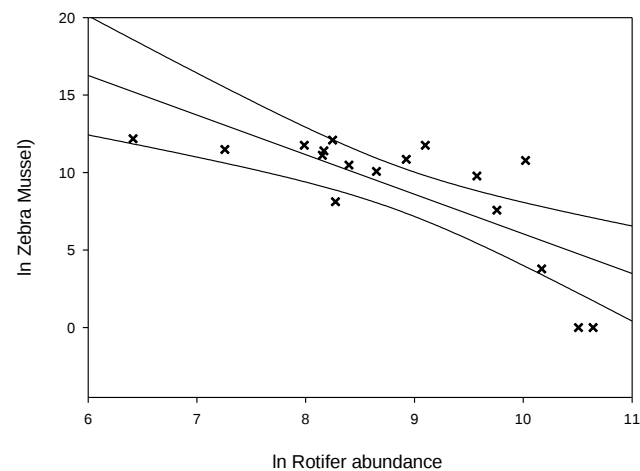
Lake Champlain long-term trends



$$r^2 = 0.56,$$

$$p = <0.001$$

Rotifer versus zebra mussel abundance Lake Champlain





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Environmental change in Lake Champlain revealed by long-term monitoring

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Dreissena

ABSTRACT

Long-term monitoring data on Lake Champlain spanning the past two to five decades were analyzed to document water quality and biological changes in the lake. August mean surface water temperatures increased during 1964–2009 in most Lake Champlain regions at rates (0.035–0.085 °C/year) similar to what has been observed in the Laurentian Great Lakes and elsewhere. Secchi disk transparency increased by over a meter during 1964–2009 in regions along the main stem of the lake, with much of the increase occurring after the 1993 zebra mussel invasion. Transparency declined in northeastern regions where zebra mussel densities were lower. No trends in hypolimnetic dissolved oxygen concentrations or depletion rates were found in any of the deep lake regions during 1990–2009. Sodium concentrations tripled in the Main Lake region since the 1960s. Chloride increased in the Main Lake by 30% since 1992, but declined in northeastern regions of the lake during recent years, coincident with reductions in road salt use in Vermont. Total phosphorus concentrations decreased during 1979–2009 in southern and northwestern lake regions, but increased by 72% in Missisquoi Bay where chlorophyll-*a* concentrations doubled over the period. There was a general lakewide trend of decreasing total nitrogen levels during 1992–2009 that may have been due in part to reductions in atmospheric nitrogen loading to the watershed. Cyanobacteria increased their dominance within the phytoplankton community in northeastern regions of the lake since the 1970s.

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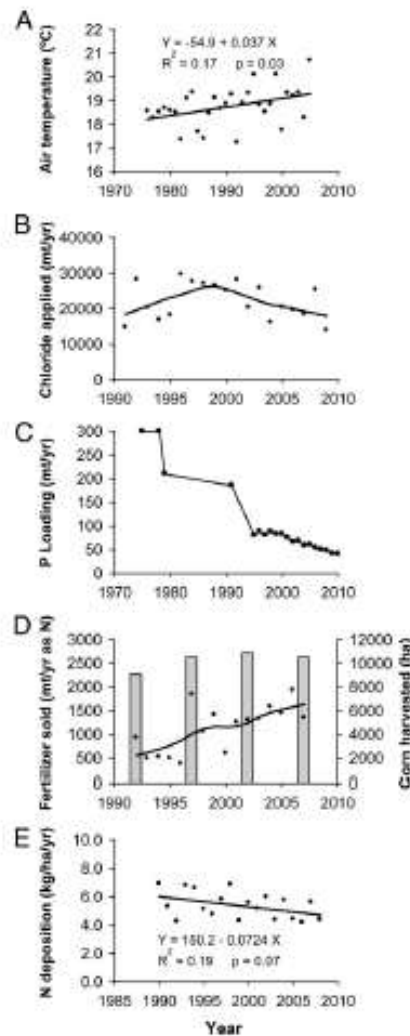
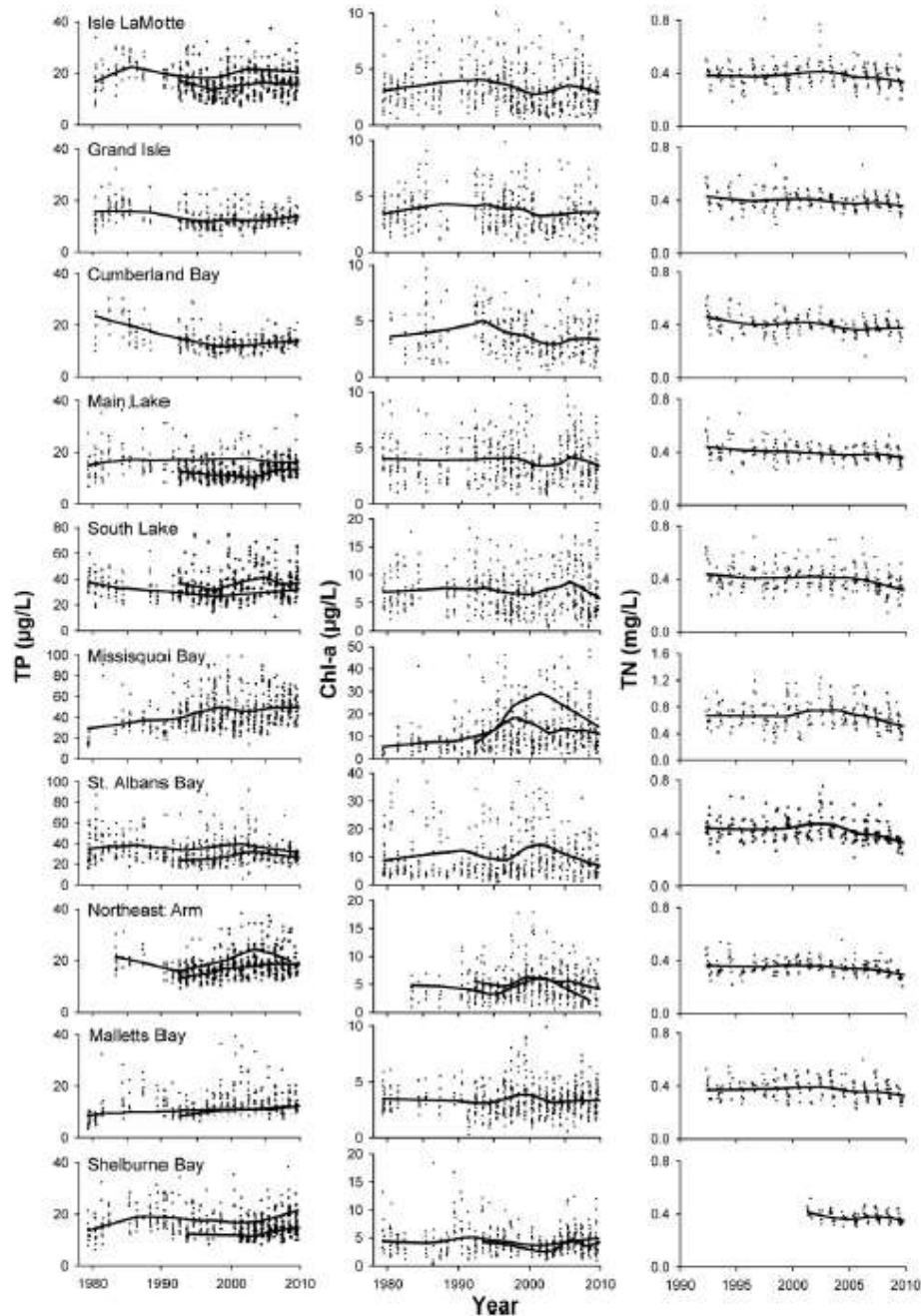


Fig. 1. Trends in Lake Champlain environmental stressors. **A.** Summer mean air temperature in the Lake Champlain Basin, 1976–2005 (modified from Stager and Thill, 2010). **B.** Winter road salt application rates (metric tons per year as chloride) for Vermont state highway maintenance districts within the Lake Champlain Basin, 1990–2009 (Vermont Agency of Transportation data). **C.** Total phosphorus loading to Lake Champlain from Vermont and New York wastewater treatment facilities, 1975–2010 (Smeltzer et al., 2009; Bogdan, 1978). **D.** Nitrogen fertilizer sold (dots and LOWESS trend line) and area of corn harvested (vertical bars) in Franklin County, VT, 1992–2007. Fertilizer sales data are from the Vermont Agency of Agriculture, Food, and Markets and do not include manure. Corn data are from the U.S. Department of Agriculture Census of Agriculture. **E.** Annual mean atmospheric wet deposition rates of inorganic nitrogen at Underhill, VT, 1990–2008 (National Atmospheric Deposition Program data).

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2012

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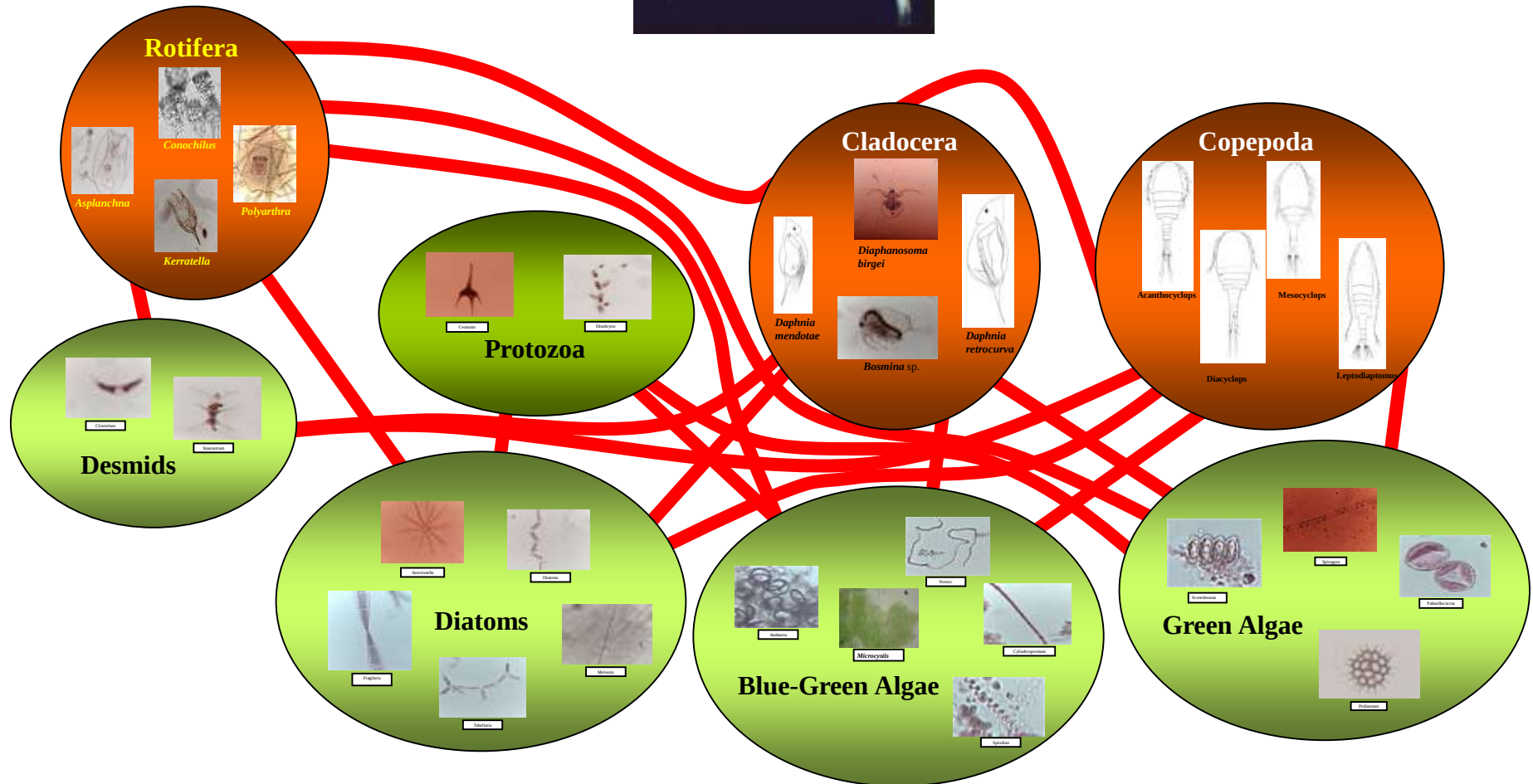


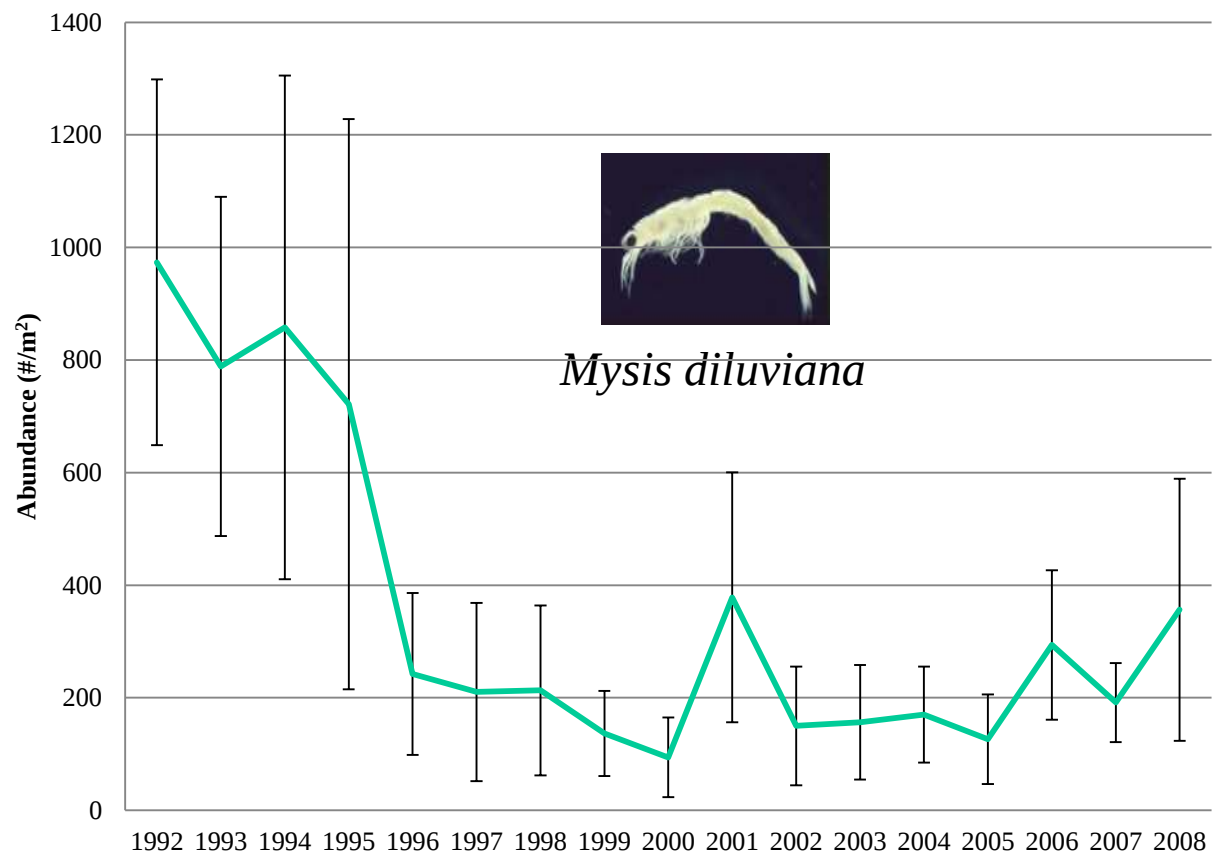
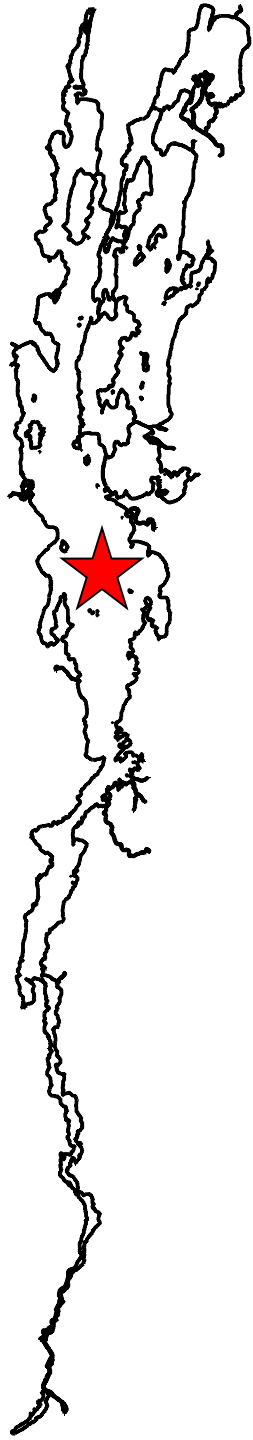
Water quality variables do not explain patterns in the Lake's plankton.



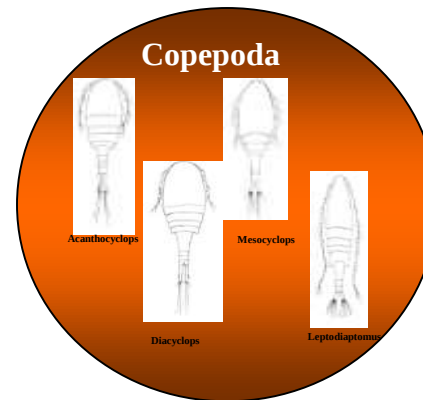
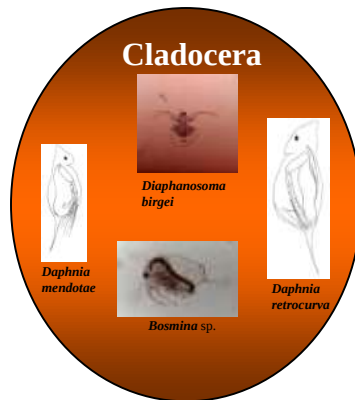
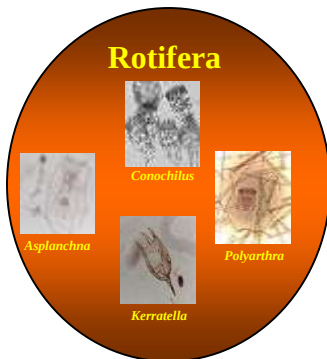
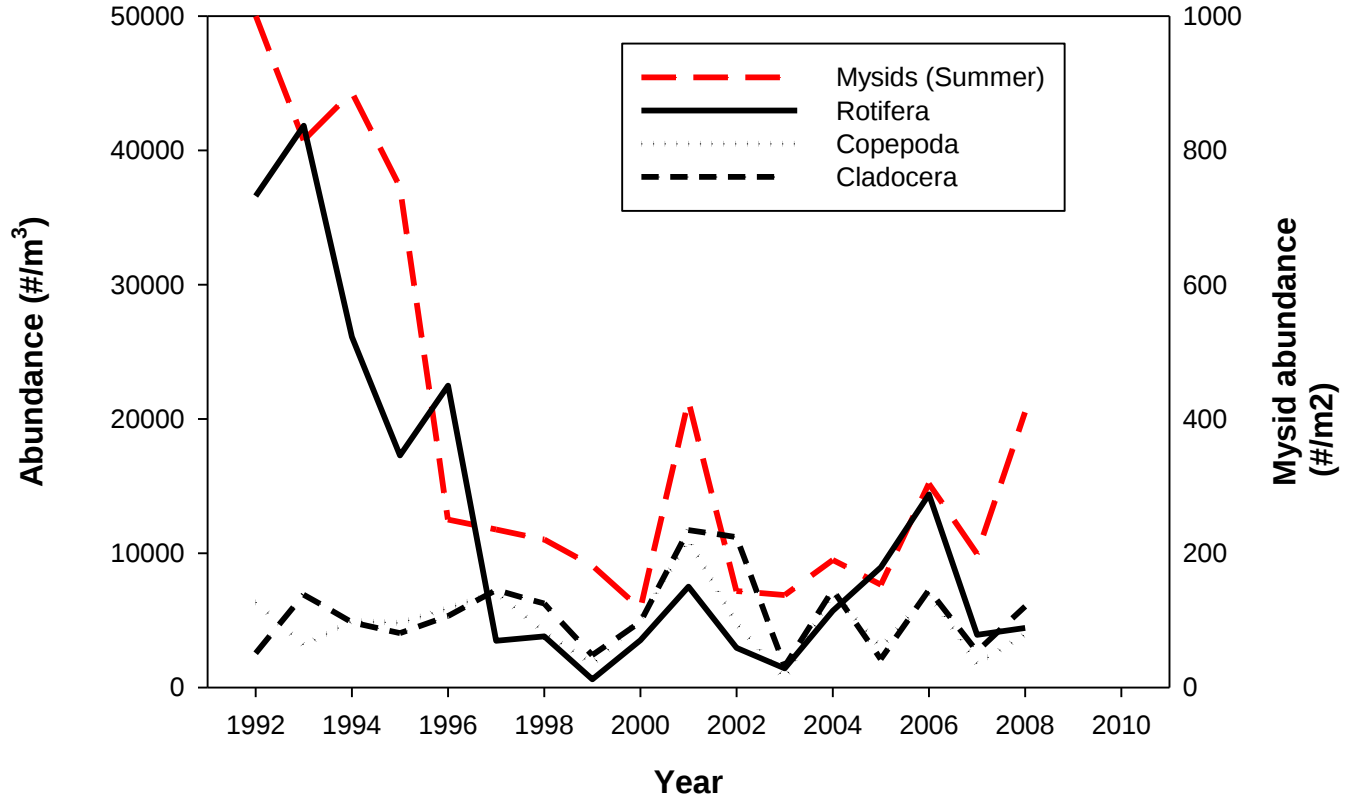


Lake Champlain Food Web

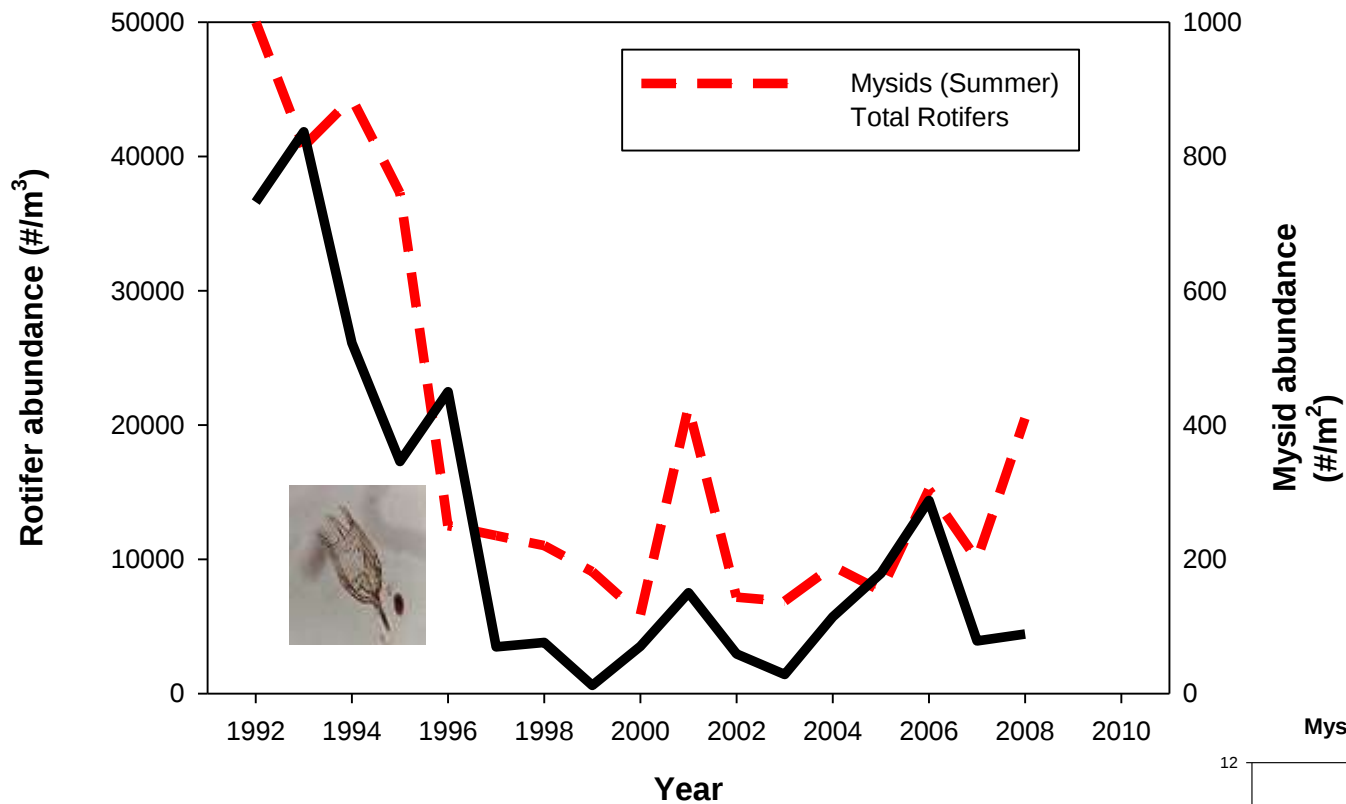




Lake Champlain Mysid long-term trends



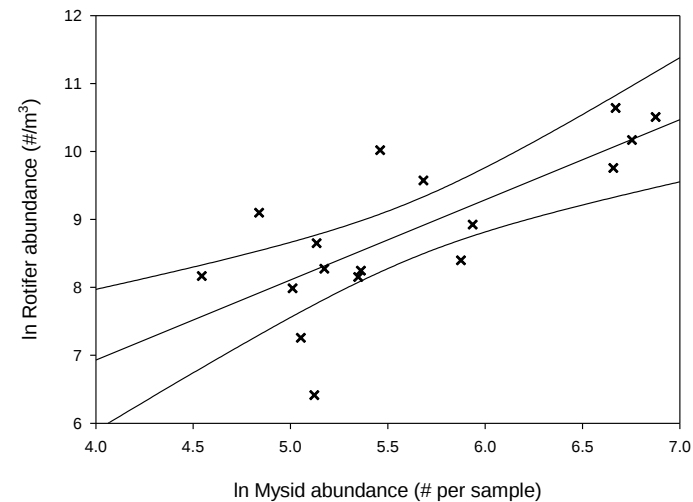
Lake Champlain Mysid long-term trends



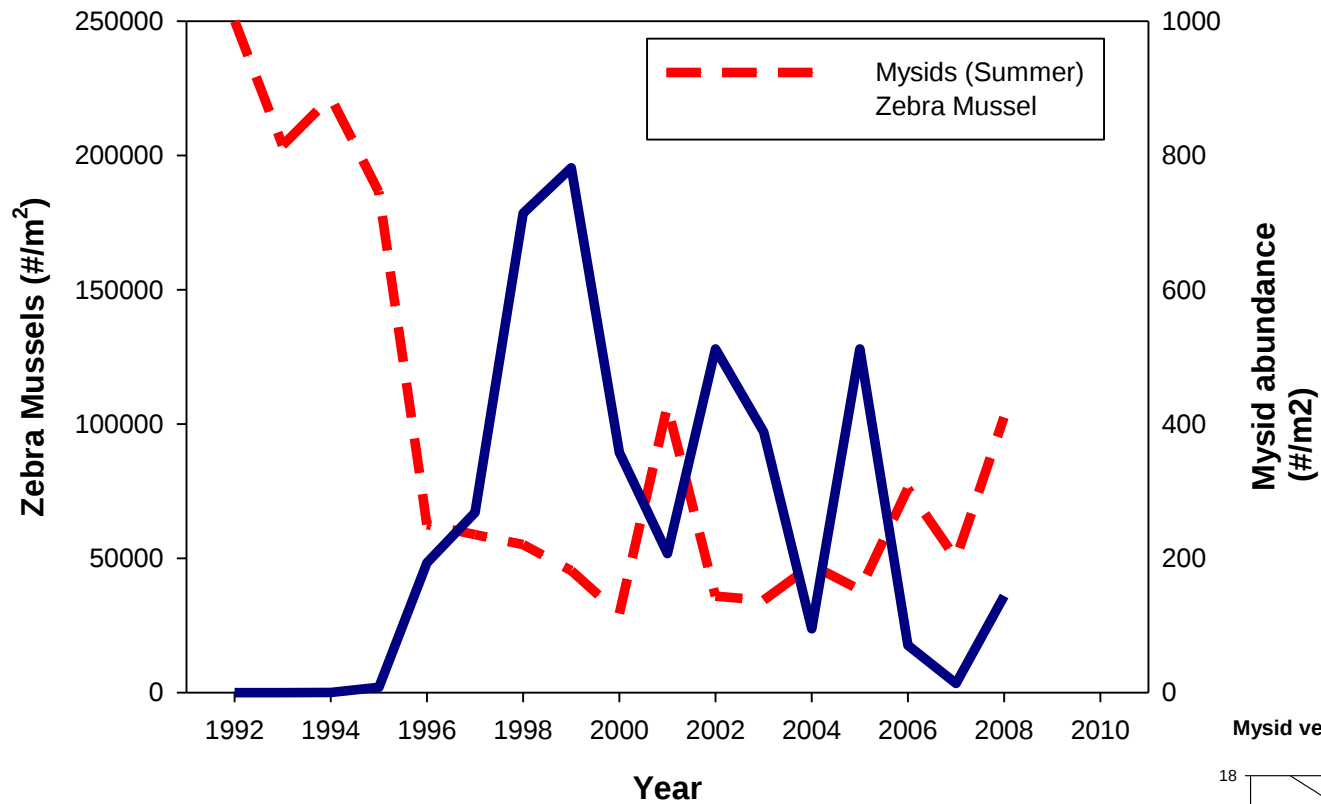
$$r^2 = 0.56,$$

$$p < 0.001$$

Mysid versus total Rotifers-Lake Champlain



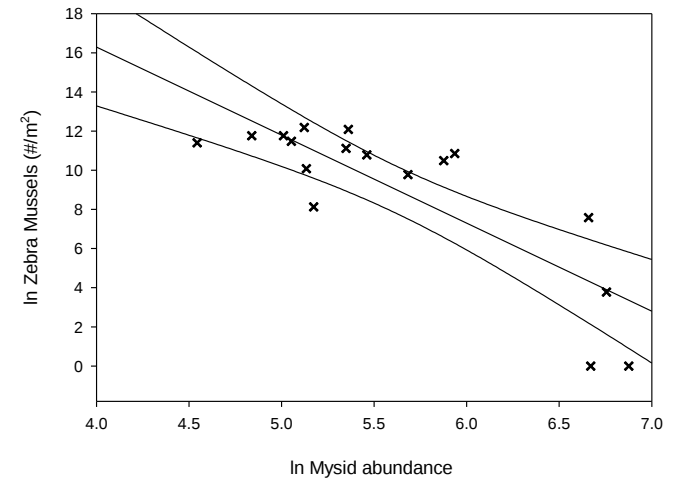
Lake Champlain Mysid long-term trends



$$r^2 = 0.68,$$

$$p = <0.001$$

Mysid versus zebra mussel abundance Lake Champlain

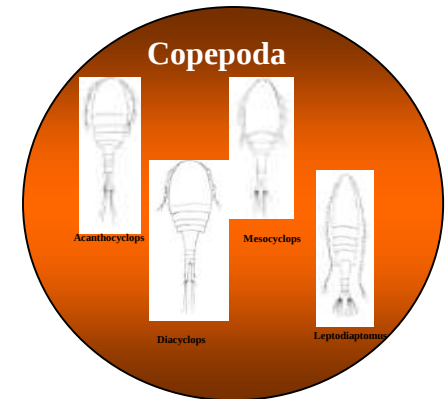
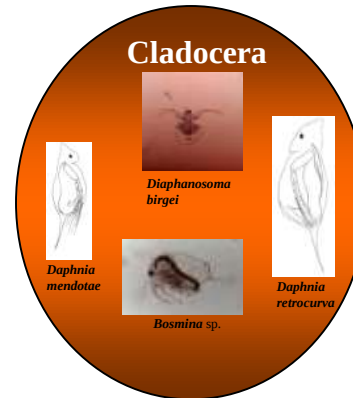




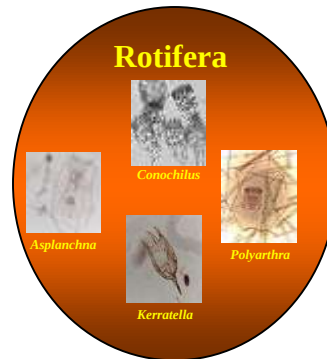
Can we model these relationships?



Trophic cascade



Filtration effect



Order of magnitude decline
in abundance

Summary

- Long-term patterns in Lake Champlain Zooplankton-

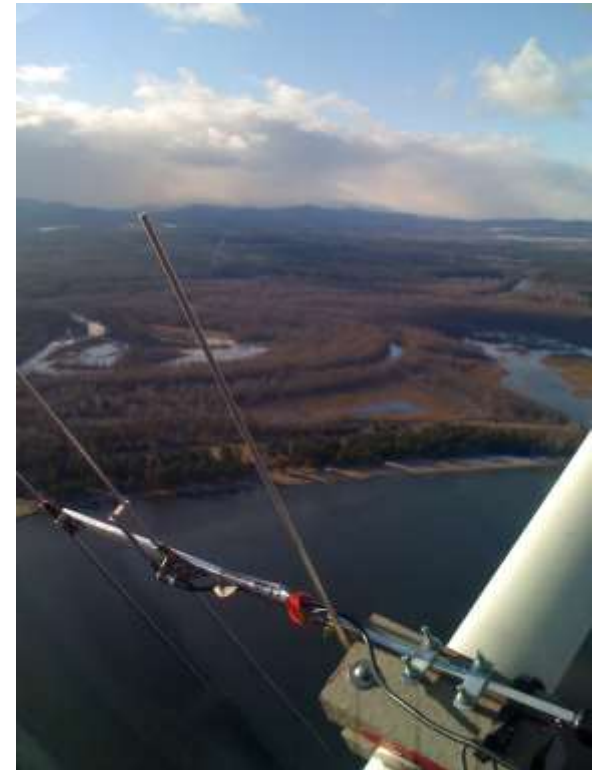
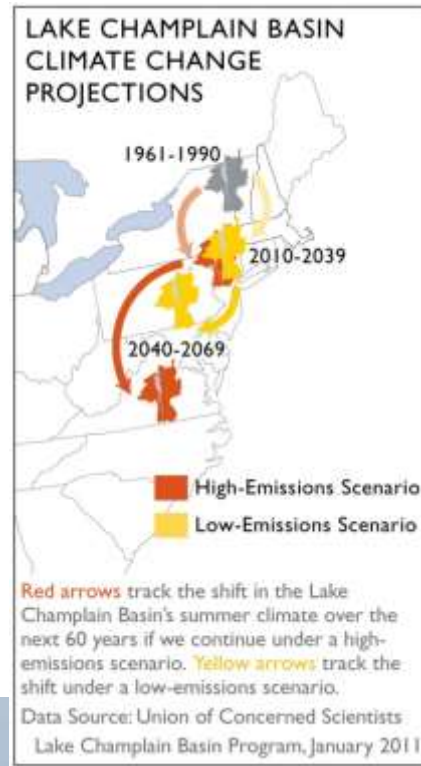
- *Zebra Mussel:*



- Rotifer declines in richness and abundance- Mid 1990s
 - Mysid declines correlate with ZM, Rotifer.... Patterns

Trophic Cascade?

Does climate change influence plankton?



Lake Champlain Ice Overs 1816-2013

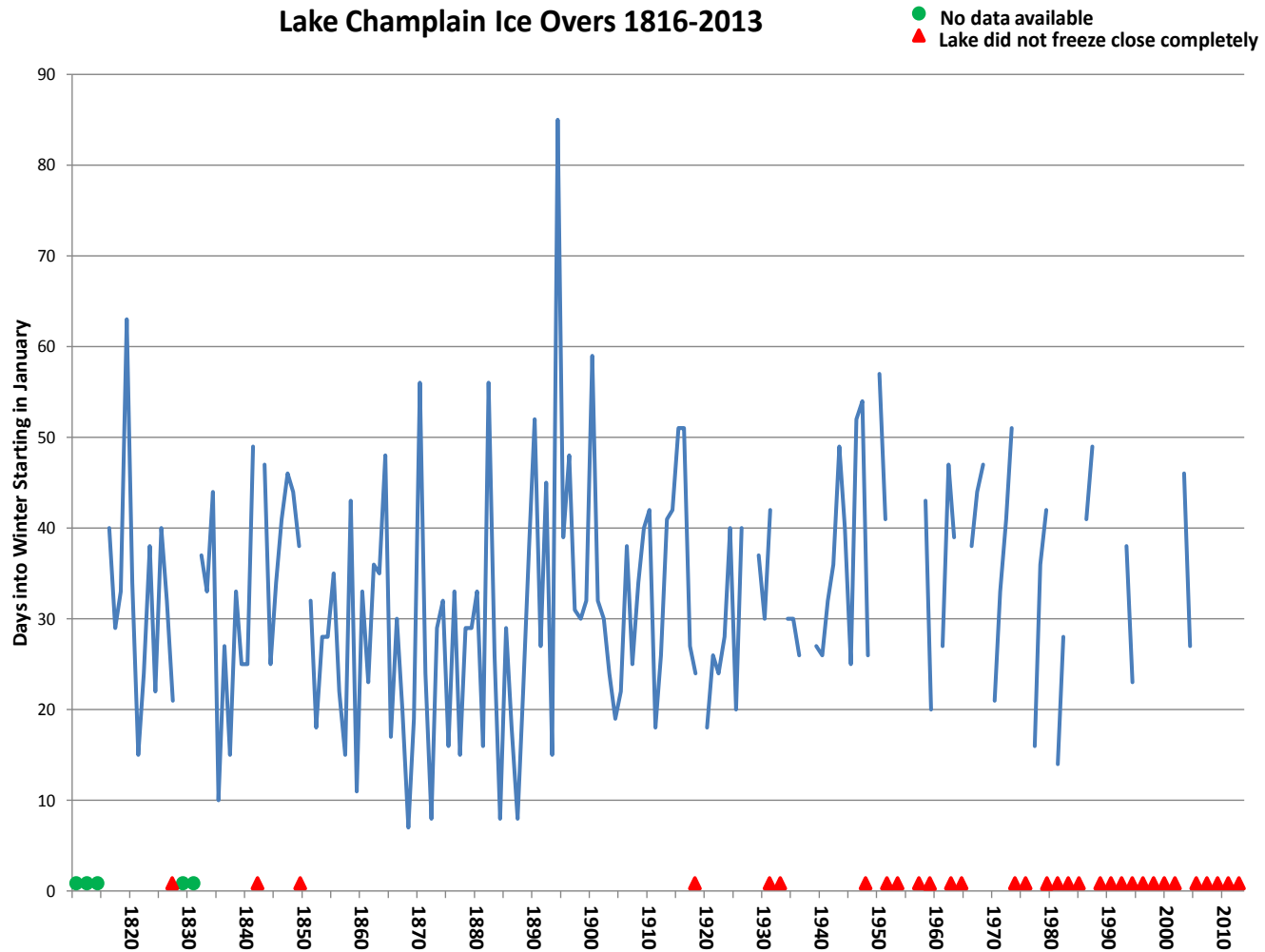


Figure 2: Lake Champlain ice over's 1816-2013. Data used in the creation of this chart was taken from NOAA.
(<http://www.erh.noaa.gov/btv/climo/lakeclose.shtml>)

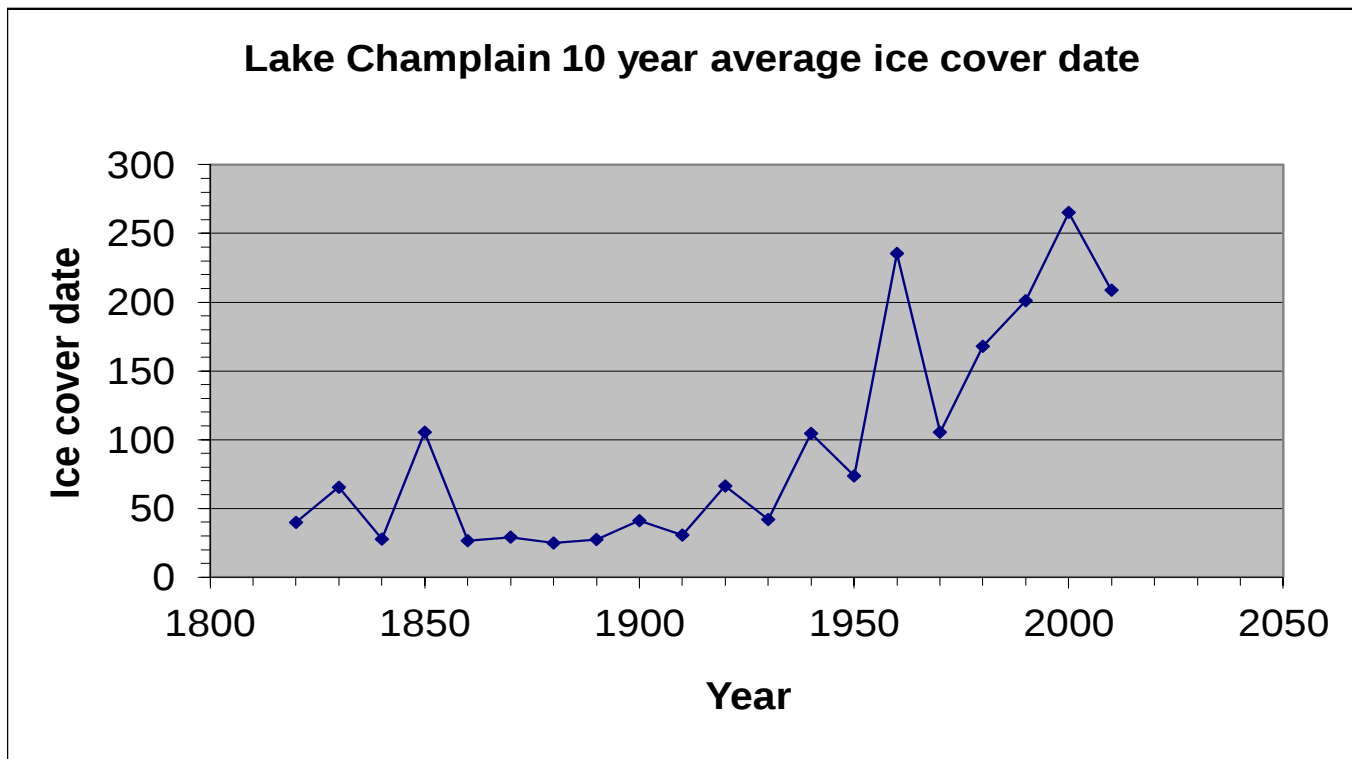
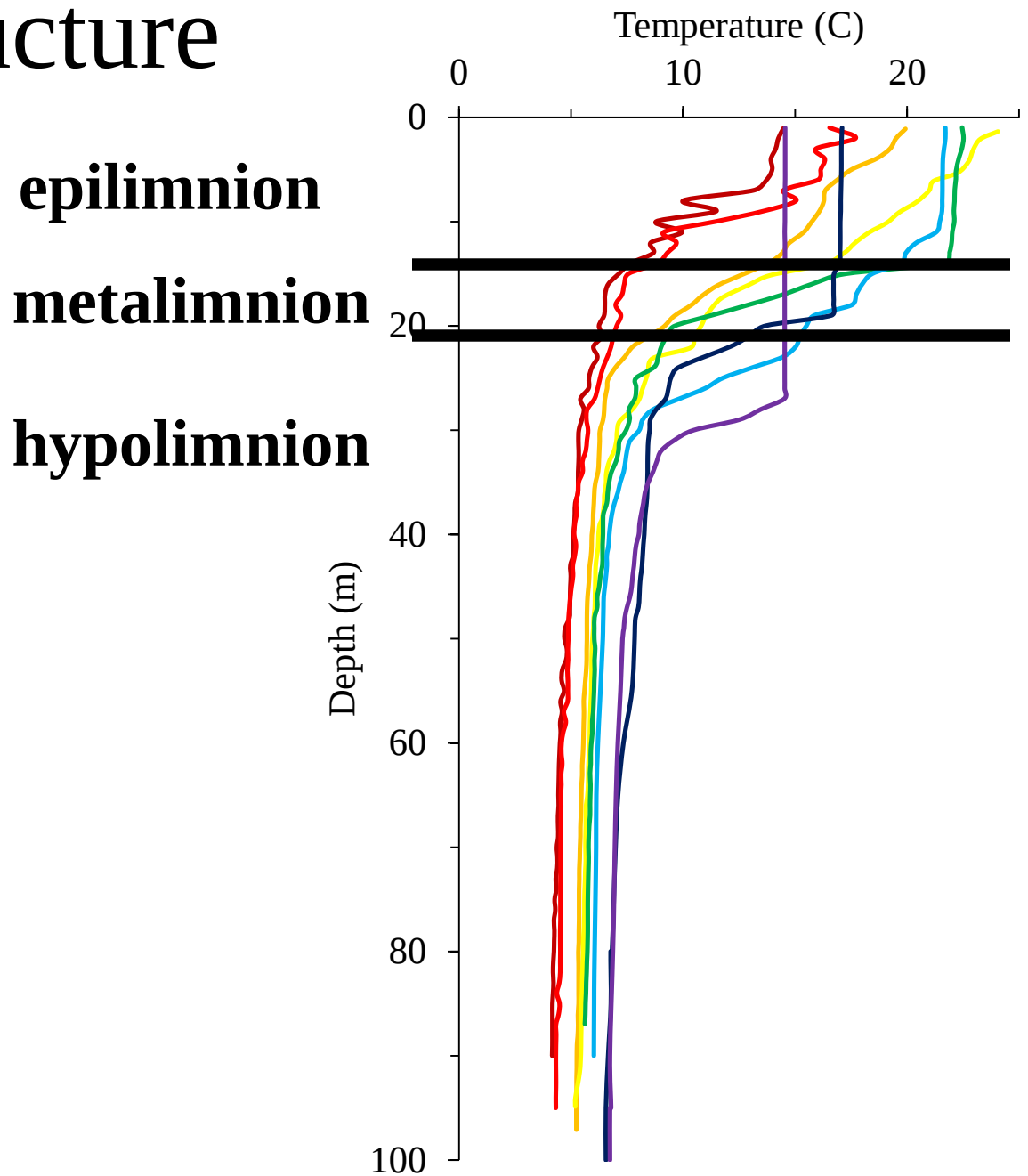
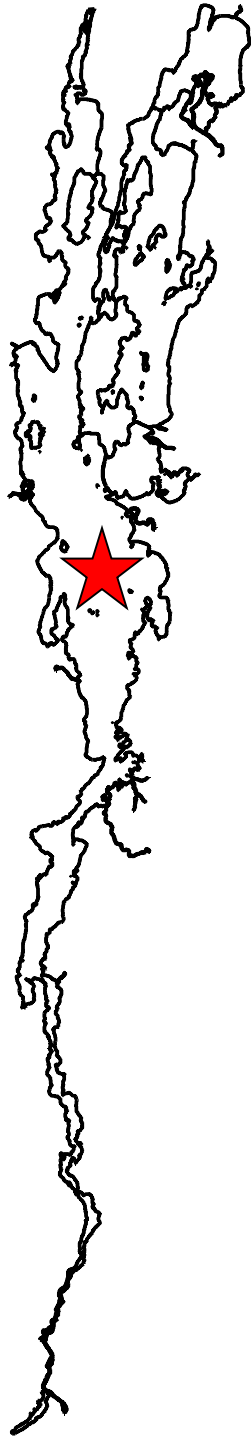


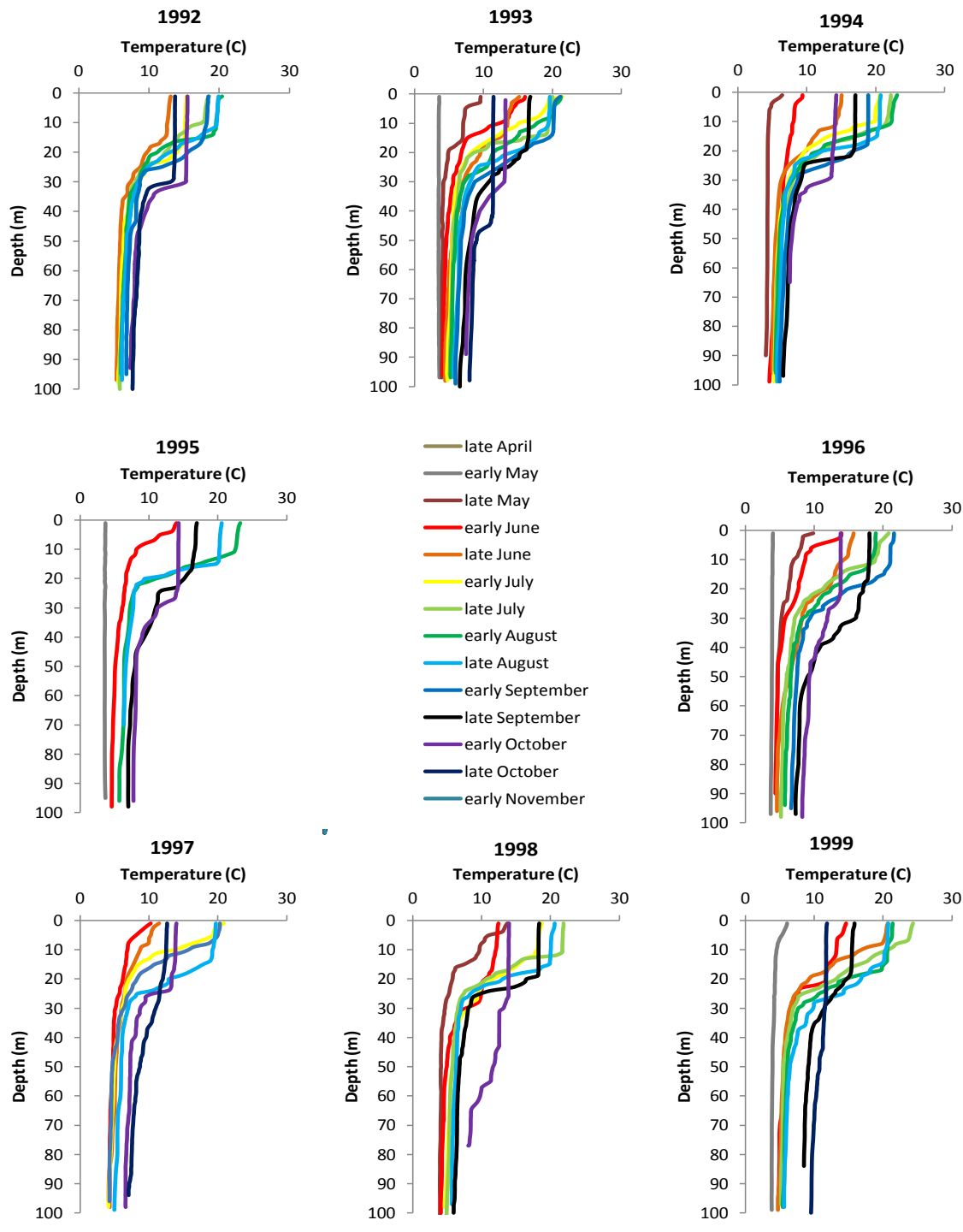
Figure 3: 10 year averages of Lake Champlain ice out data from 1816-2010. Ice out data obtained from NOAA (<http://www.weather.gov/btv/lakeclose>).

Thermal structure





Station 19



3 Year Average Maximum Epilimnion Depth Station 19

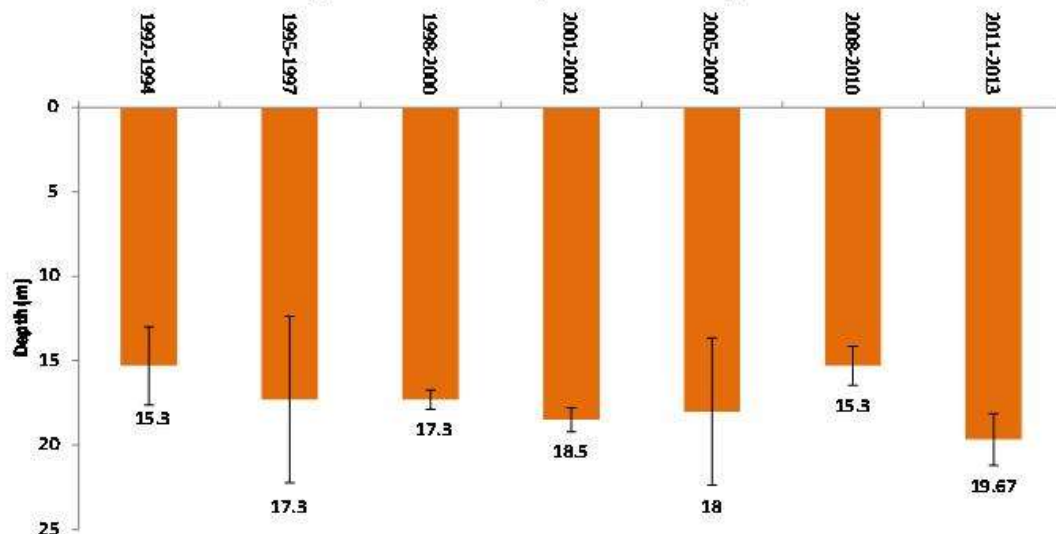


Figure 11: 3 year averages of maximum epilimnion depth at Station 19 from 1992-2013. 2003-2004 data are missing.

3 Year Average Minimum Hypolimnion Depth station 19

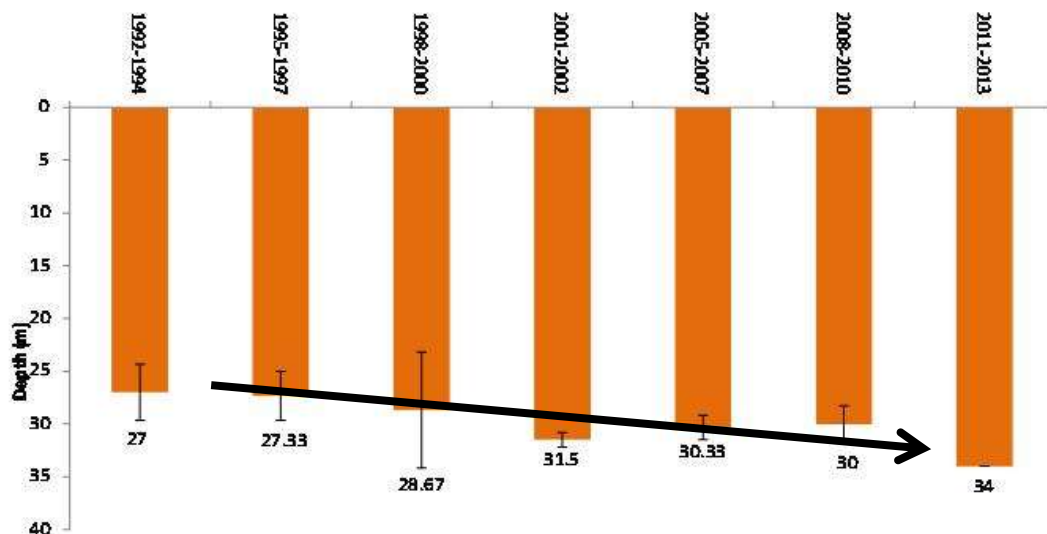
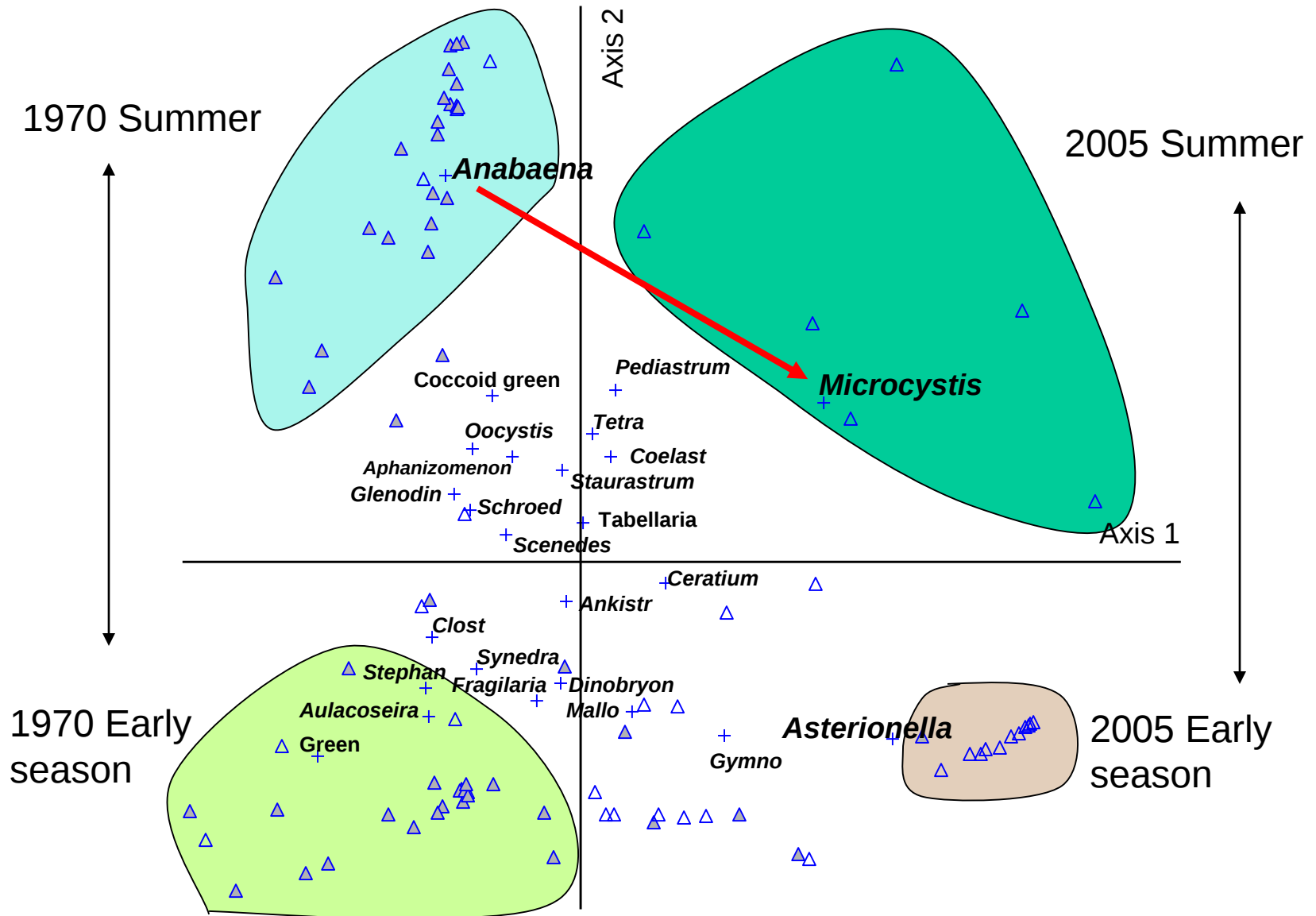


Figure 12: 3 year averages of minimum hypolimnion depth at Station 19 from 1992-2013. 2003-2004 data are missing.

**An In-Depth Comparison of Lake Champlain
Phytoplankton Community Composition and
Distribution from 1970 to 2005**

Shelley L. Bouyea

1970 vs 2005 phytoplankton NMS results



Timothy B. Mihuc, Carrienne Pershyn, Sean Thomas, Greg Boyer, Mike Satchwell, Jeffry Jones, and Meghan Greene

Timothy B. Mihuc, Carrienne Pershyn, Sean Thomas, Greg Boyer, Mike Satchwell, Jeffry Jones, and Meghan Greene

July 27

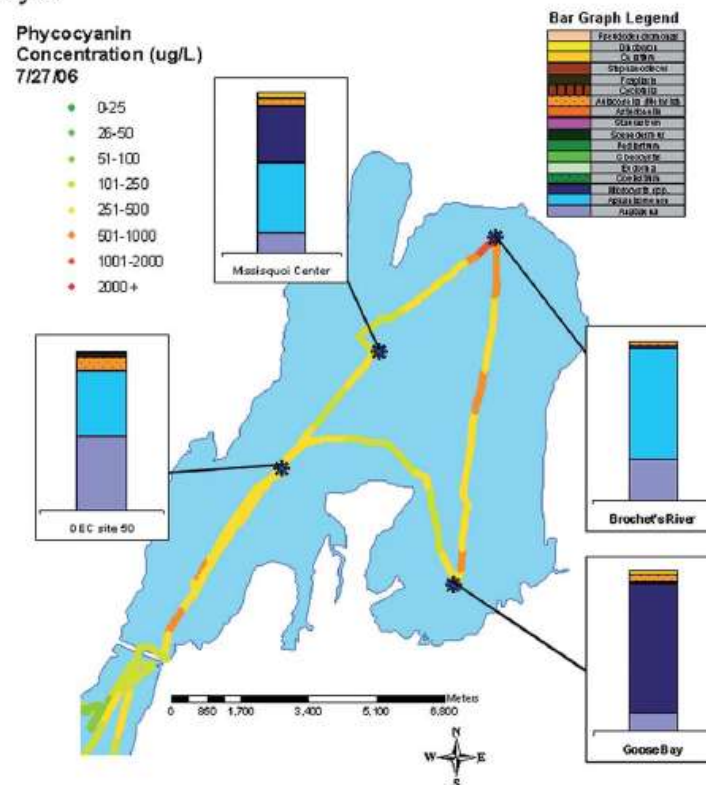
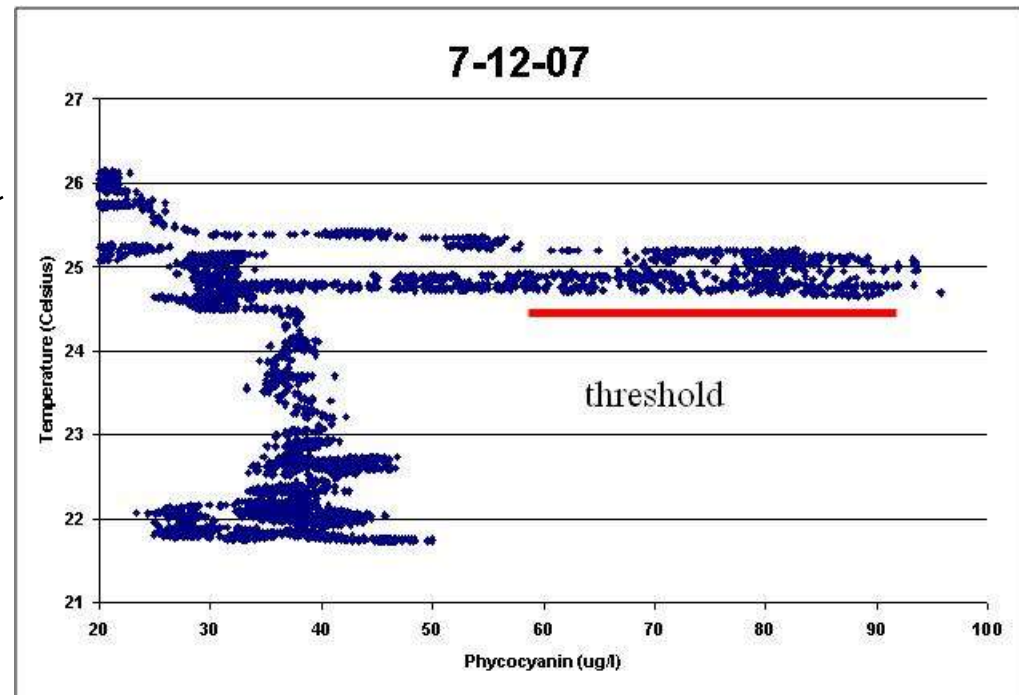
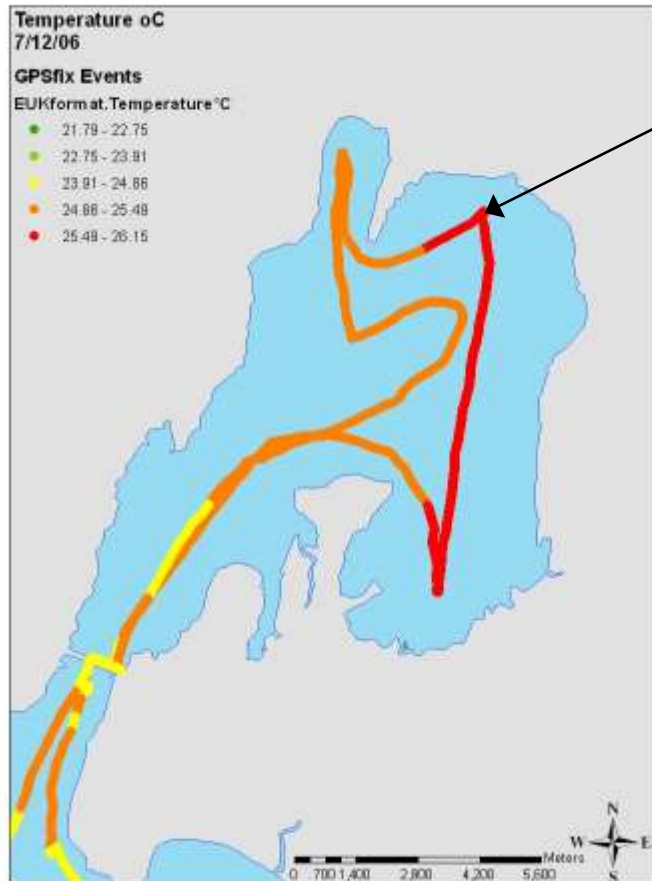
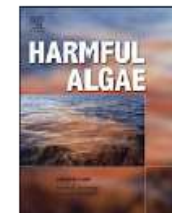


Fig. 2. Bloom map of Missisquoi Bay, Lake Champlain, for 27 July 2006. Phycocyanin concentration ($\mu\text{g/L}$) is shown along the mapping track with phytoplankton community composition (% composition) at selected locations in the inset graphs. Bar graph legend indicates (from top to bottom) protozoa (entries 1–3), diatoms (entries 4–8; brown-yellow shading), desmids (*Staurastrum*, pink), green algae (entries 10–14; green shades), and Cyanobacteria (last 3 entries; blue shading).



- Species composition for all sites in Missisquoi Bay was dominated by diatoms, namely *Aulacoseira*.
- *Microcystis spp.* and *Aphanizomenon spp.* were present in low abundance at Venise Bay, QC and Brochet's River, QC, which may be attributed to the higher water temperatures in that part of the bay on this date
- A temperature threshold may exist for large bloom formation.



The effects of temperature and nutrients on the growth and dynamics of toxic and non-toxic strains of *Microcystis* during cyanobacteria blooms

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Microcystin synthetase

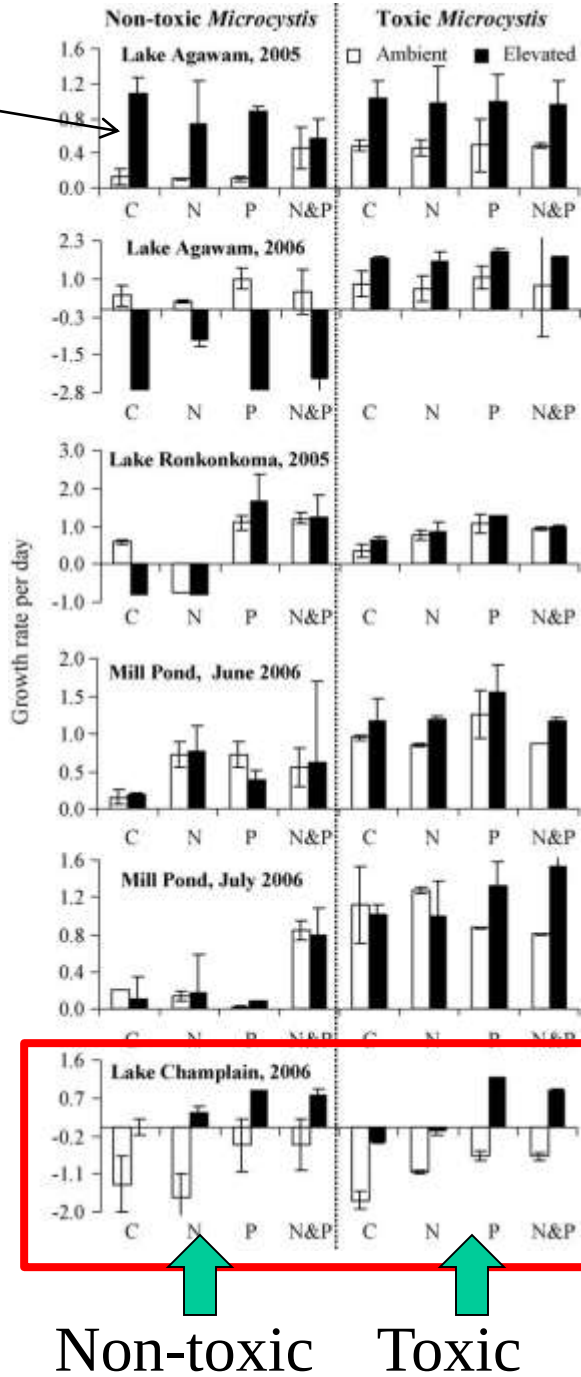
Harmful algal bloom

ABSTRACT

In temperate latitudes, toxic cyanobacteria blooms often occur in eutrophied ecosystems during warm months. Many common bloom-forming cyanobacteria have toxic and non-toxic strains which co-occur and are visually indistinguishable but can be quantified molecularly. Toxic *Microcystis* cells possess a suite of microcystin synthesis genes (*mcyA–mcyJ*), while non-toxic strains do not. For this study, we assessed the temporal dynamics of toxic and non-toxic strains of *Microcystis* by quantifying the microcystin synthetase gene (*mcyD*) and the small subunit ribosomal RNA gene, 16S (an indicator of total *Microcystis*), from samples collected from four lakes across the Northeast US over a two-year period. Nutrient concentrations and water quality were measured and experiments were conducted which examined the effects of elevated levels of temperatures (+4 °C), nitrogen, and phosphorus on the growth rates of toxic and non-toxic strains of *Microcystis*. During the study, toxic *Microcystis* cells comprised between 12% and 100% of the total *Microcystis* population in Lake Ronkonkoma, NY, and between 0.01% and 6% in three other systems. In all lakes, molecular quantification of toxic (*mcyD*-possessing) *Microcystis* was a better predictor of *in situ* microcystin levels than total cyanobacteria, total *Microcystis*, chlorophyll *a*, or other factors, being significantly correlated with the toxin in every lake studied. Experimentally enhanced temperatures yielded significantly increased growth rates of toxic *Microcystis* in 83% of experiments conducted, but did so for non-toxic *Microcystis* in only 33% of experiments, suggesting that elevated temperatures yield more toxic *Microcystis* cells and/or cells with more *mcyD* copies per cell, with either scenario potentially yielding more toxic blooms. Furthermore, concurrent increases in temperature and P concentrations yielded the highest growth rates of toxic *Microcystis* cells in most experiments suggesting that future eutrophication and climatic warming may additively promote the growth of toxic, rather than non-toxic, populations of *Microcystis*, leading to blooms with higher microcystin content.

Elevated temperature

Fig. 6. Net growth rates of toxic Microcystis (right half of figure) and nontoxic Microcystis (left half of figure) during nutrient amendment experiments (t = 72 h) conducted in various systems during the 2005 and 2006 field seasons at ambient (white bars) and elevated (black bars) temperatures. C: control; N: nitrate; P: rthophosphate. Error bars represent 1 SD of triplicate experimental bottles.



“During the Lake Champlain experiment, ambient water temperatures were 24.9 2.0 8C. Experimentally enhanced temperatures (29.1 1.3 8C) significantly increased the growth rates of both toxic and non-toxic Microcystis populations by 80% and 101% respectively (p < 0.001; Table 3; Fig. 6).”

“Finally, nutrients (N or P) interacted with temperature, to enhance toxic Microcystis growth rates (p < 0.01; Table 3; Fig. 6) with the enhanced P and temperature treatment yielding the highest growth rates of any population among all treatments (1.17 0.03 d1).”

What do we know?

- Invasive species (Zebra mussel) impact planktonic communities
- Some trends (shifts in phytoplankton) may be linked to climate change.
- Water quality variables do not explain long-term patterns in the Lake's zooplankton

Lake Champlain zooplankton community dynamics following an extreme flood event



Erin Hayes-Pontius
M.S. Natural Science
research thesis
SUNY Plattsburgh

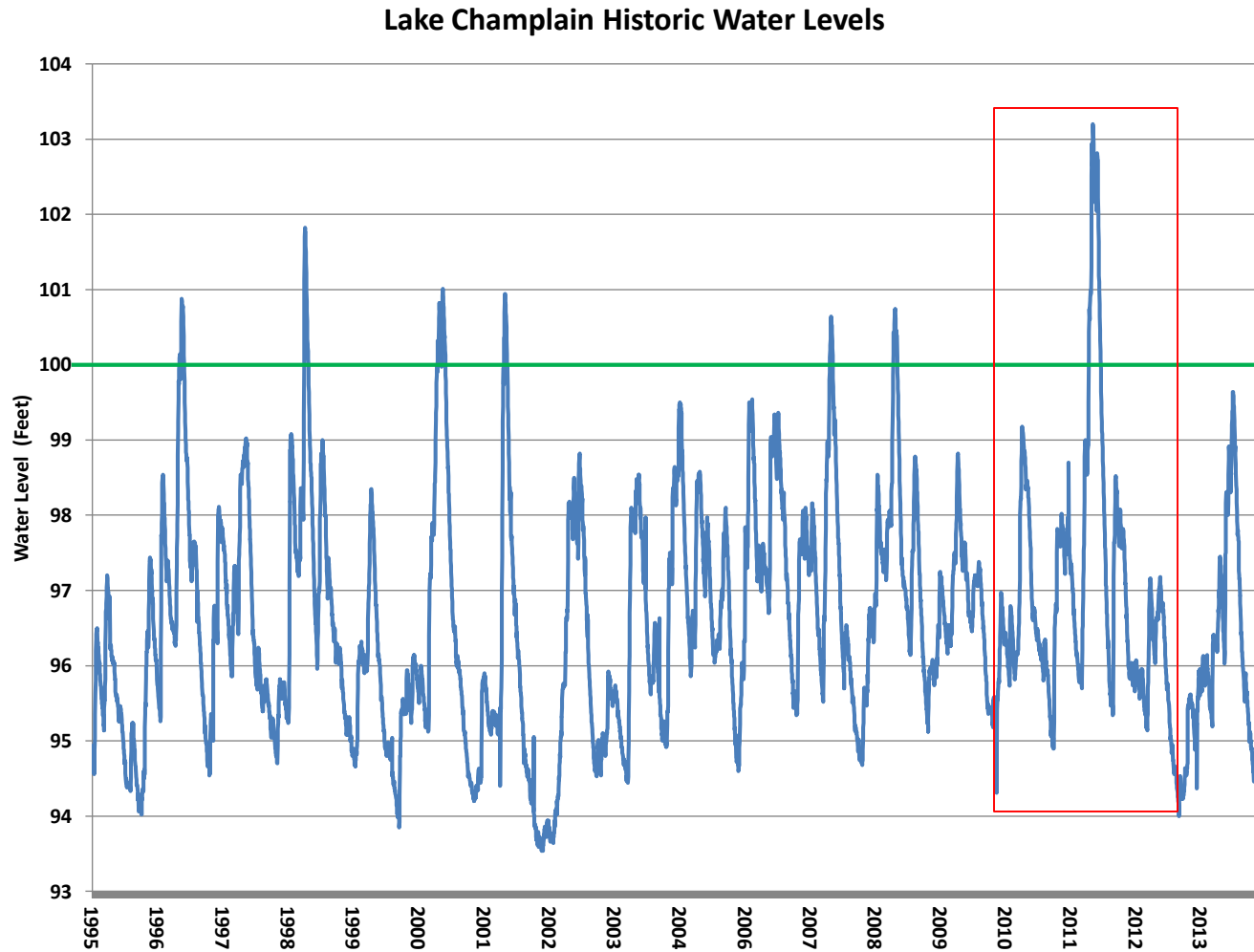
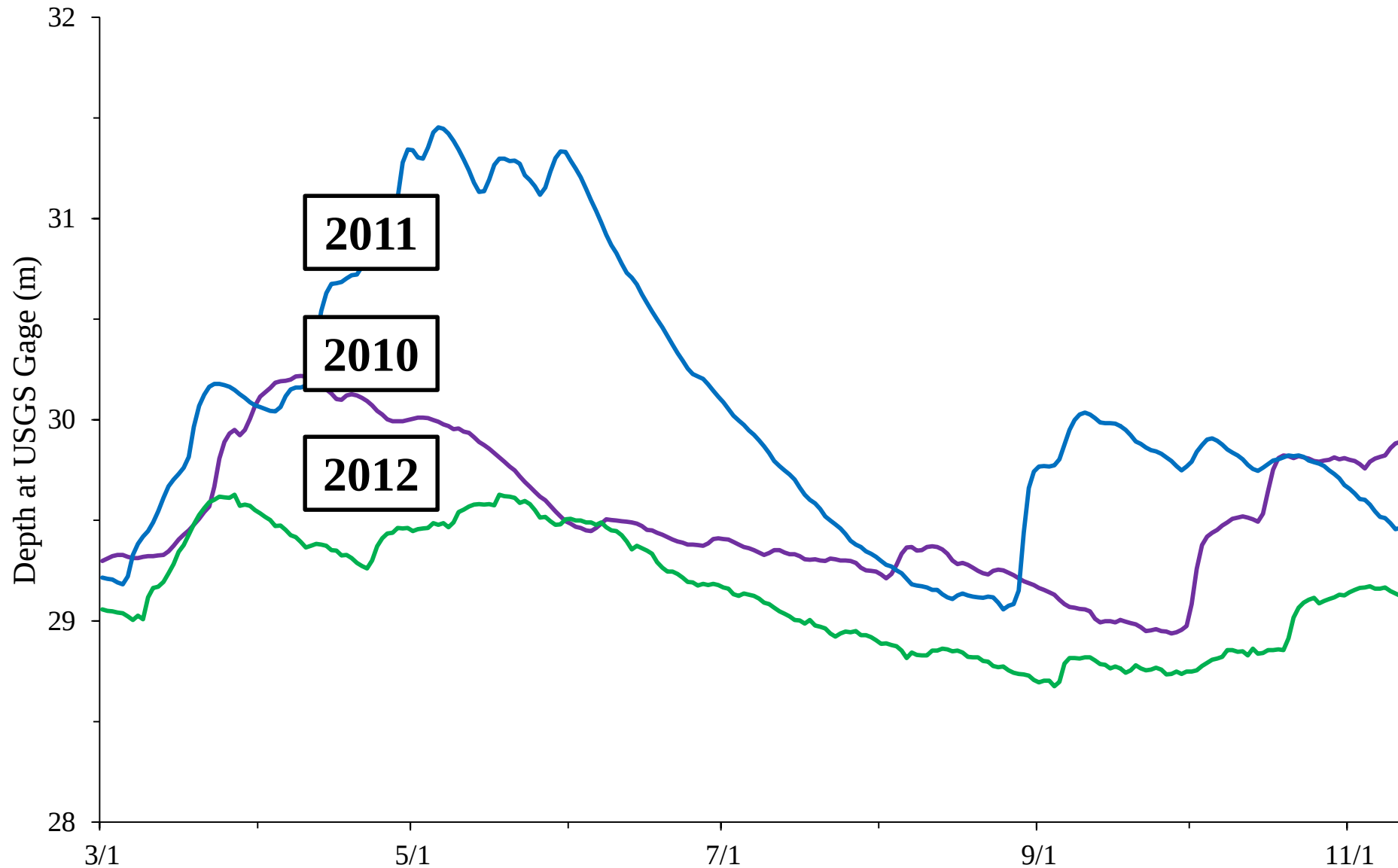


Figure 1: Lake Champlain historic water levels 1995-2013. The green line represents the national weather service flood level. Data used in the creation of this chart was taken from NOAA. (<http://www.erh.noaa.gov/btv/html/lake.php>)

Intro- lake depth



Sediment plumes



These photos are from the Lake Champlain Basin Program

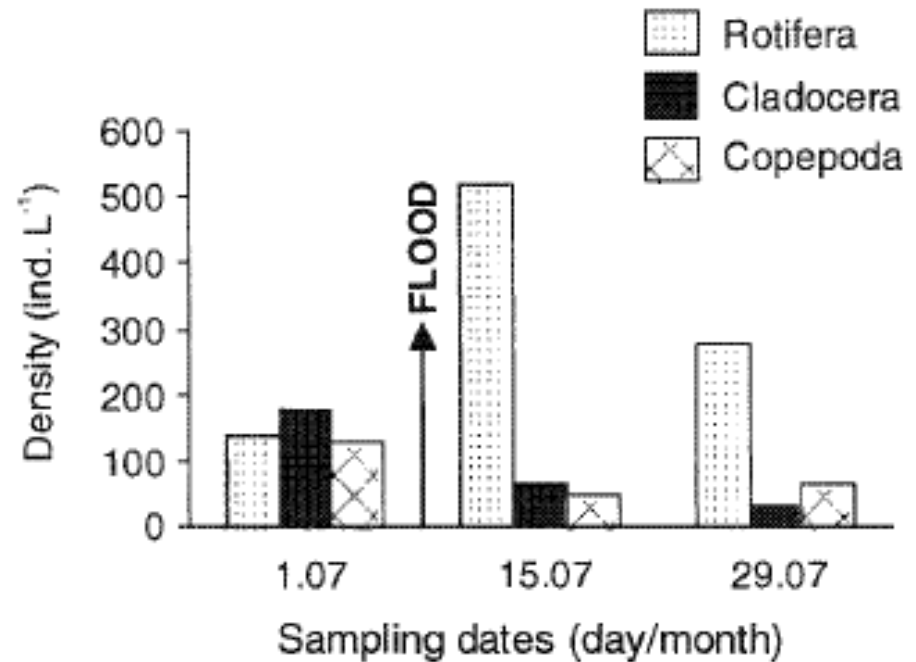
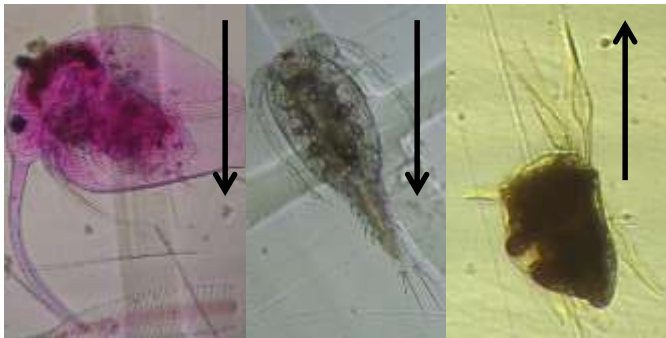
The flooding

- Lack of literature from large lakes
- What happens?



Flood responses

- Overall decreases in density
- Some groups may benefit

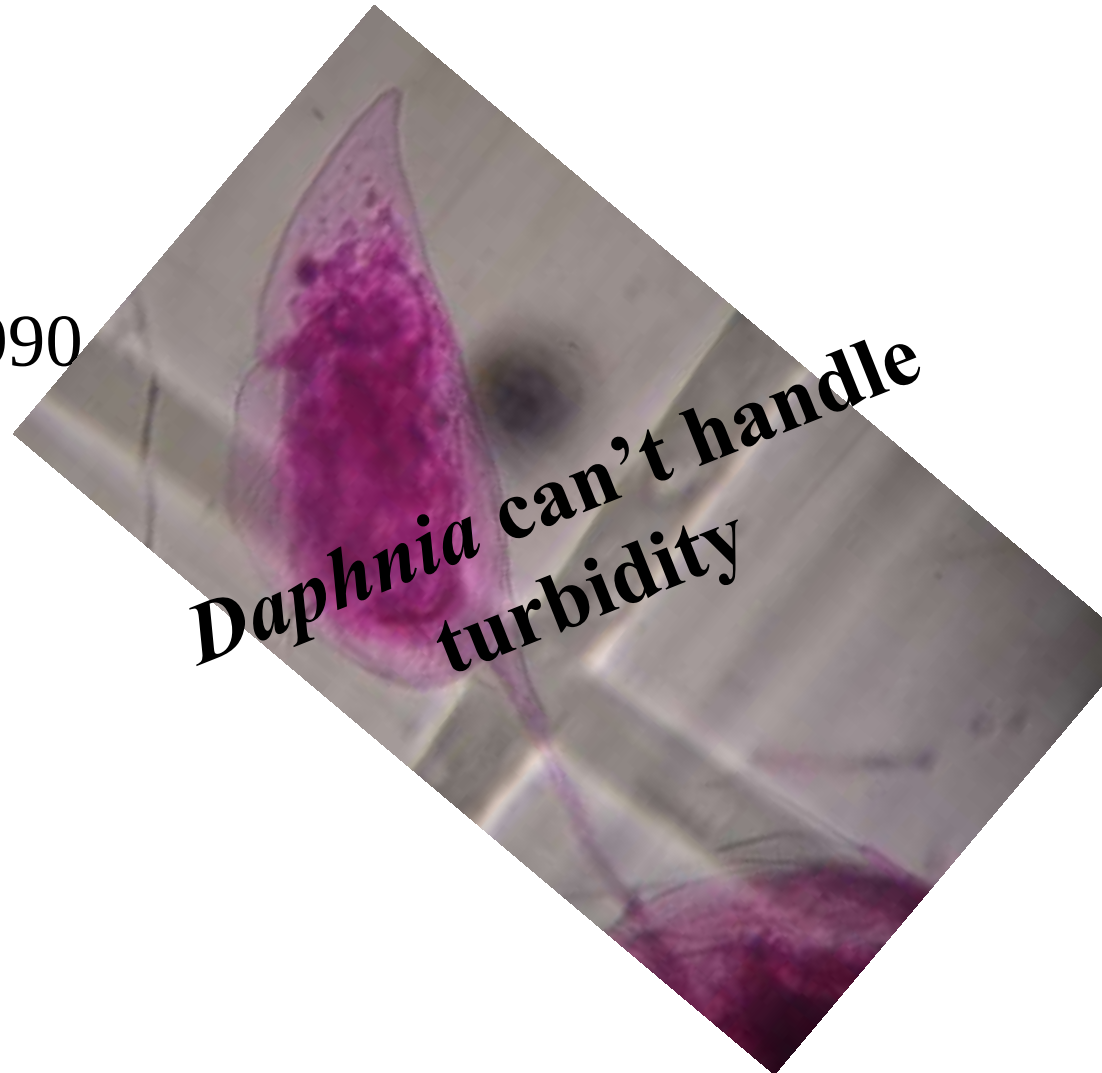


Godlewska et al.
2003

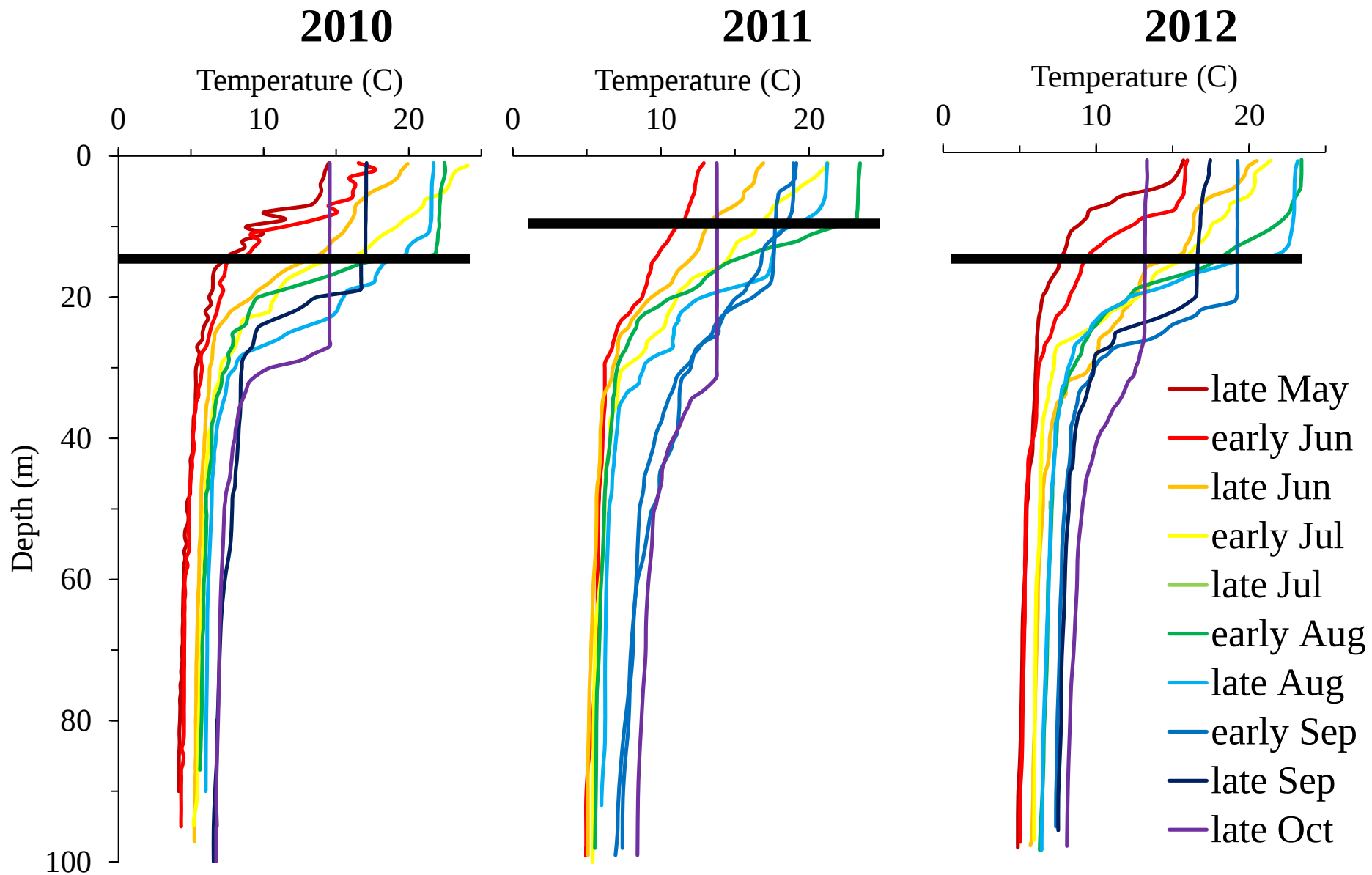
-
- This collage displays 15 individual microscopic images of mites, arranged in a grid-like fashion. The images show a variety of mite species, characterized by their small, oval or elongated bodies, segmented legs, and internal anatomical features. Some mites have prominent dark spots or structures on their bodies, while others show more complex internal patterns. The background of the images is a light, slightly textured surface, typical of a microscope slide.

Flood responses

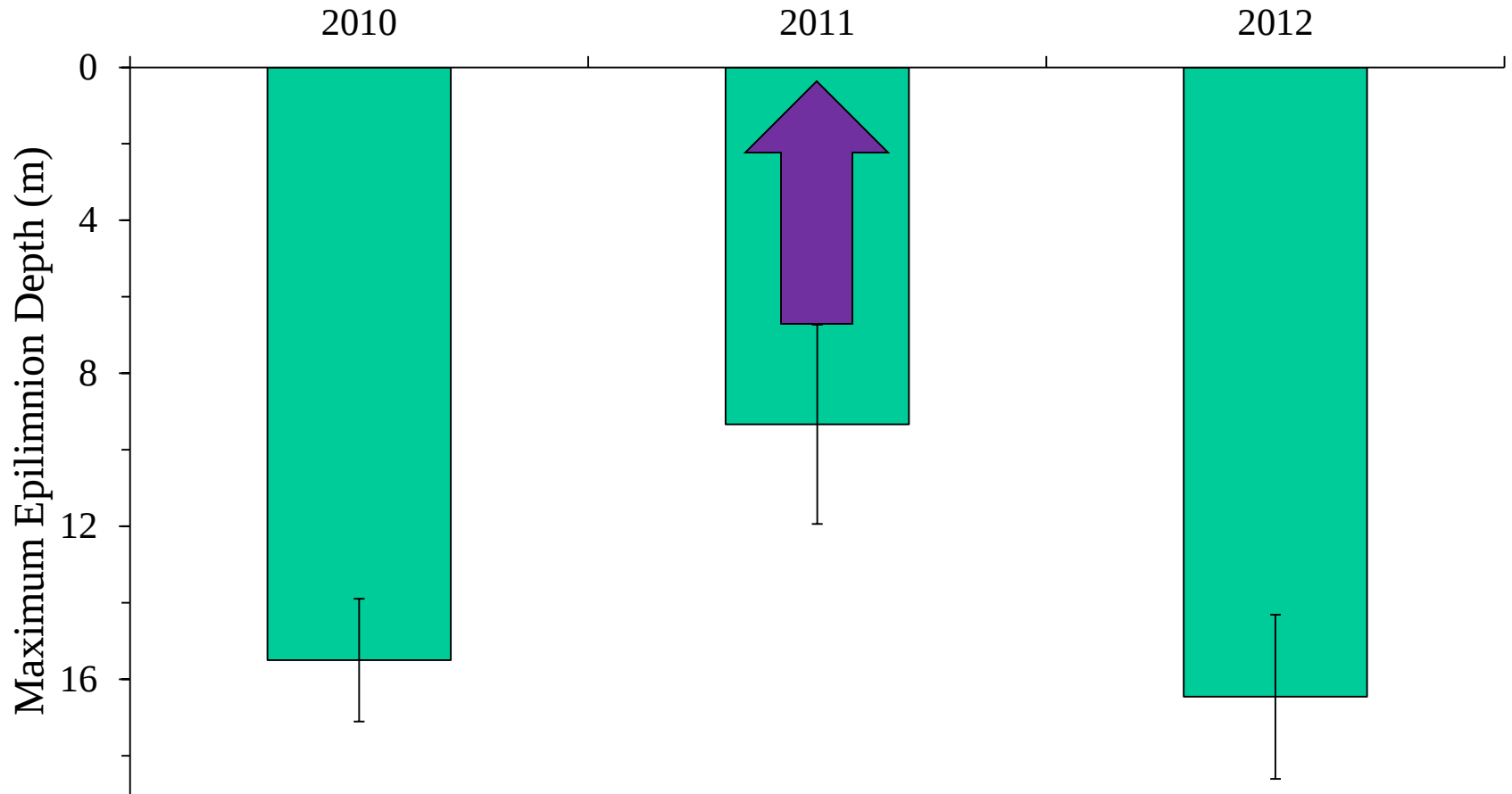
- Everyone agrees...
 - Arruda et al. 1983
 - Hart 1986
 - Threlkeld 1986
 - Kirk and Gilbert 1990
 - Dejen et al. 2004
 - Schou et al. 2009



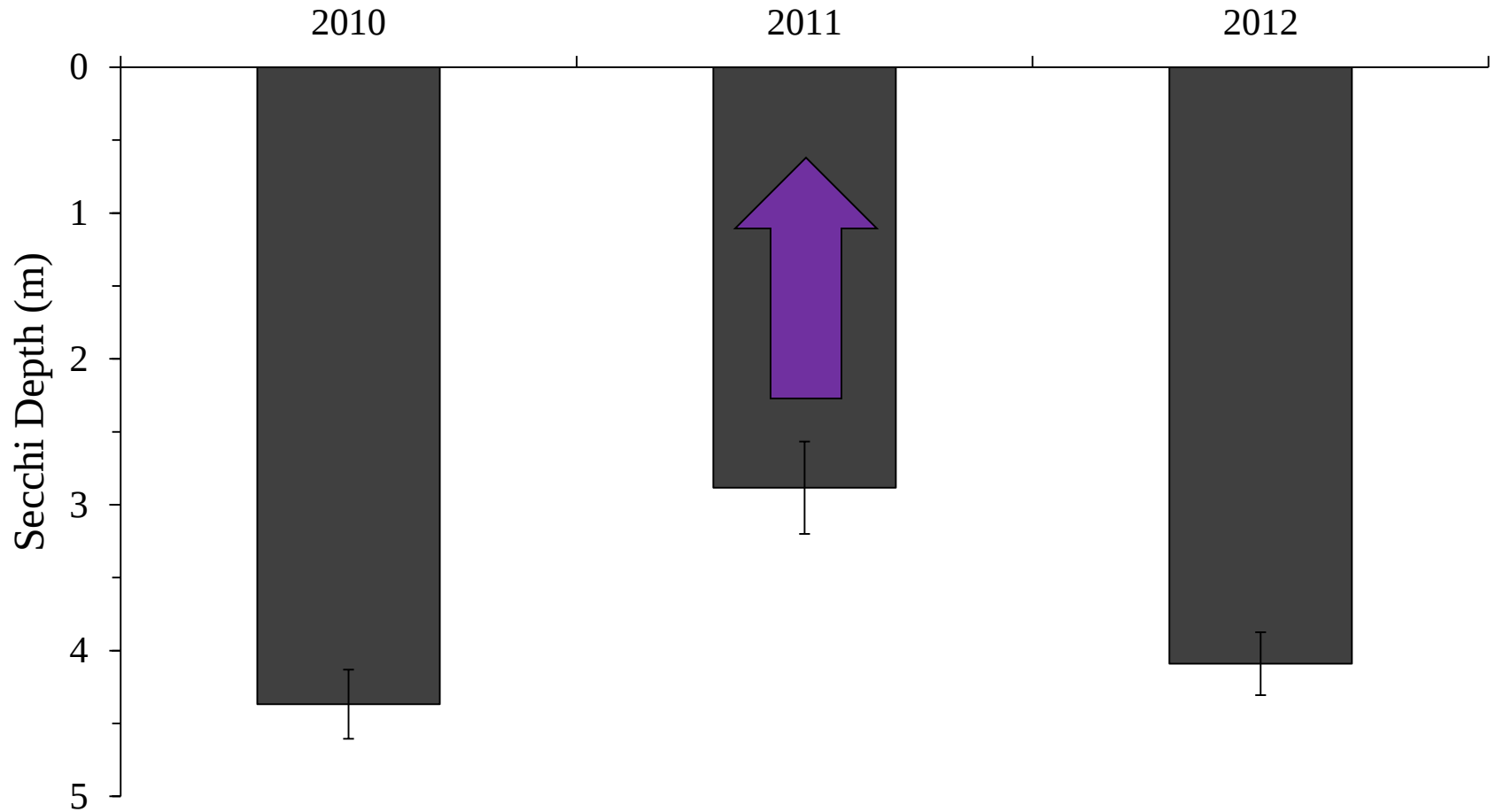
Results- vertical profiles



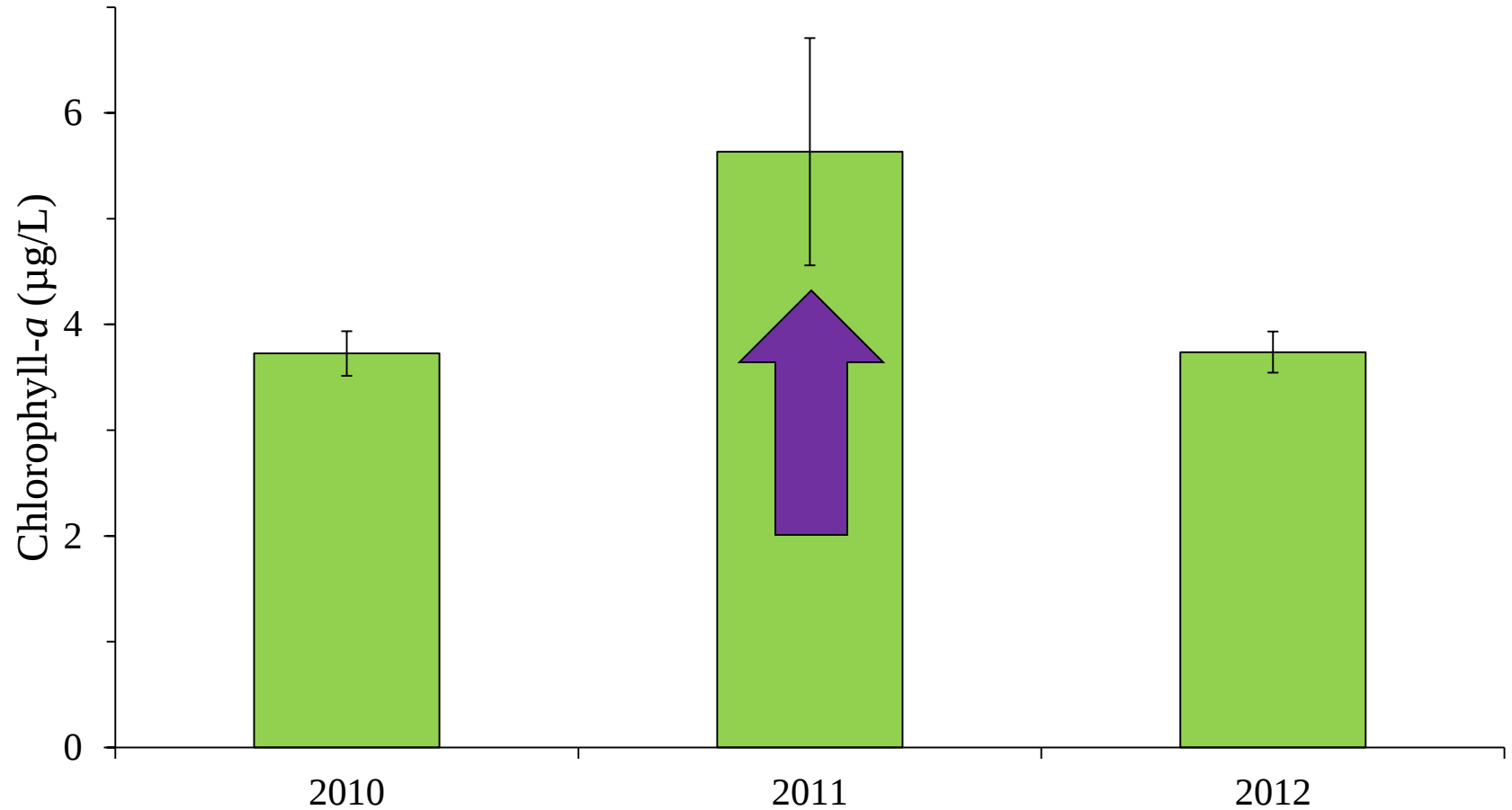
Results- thermal structure



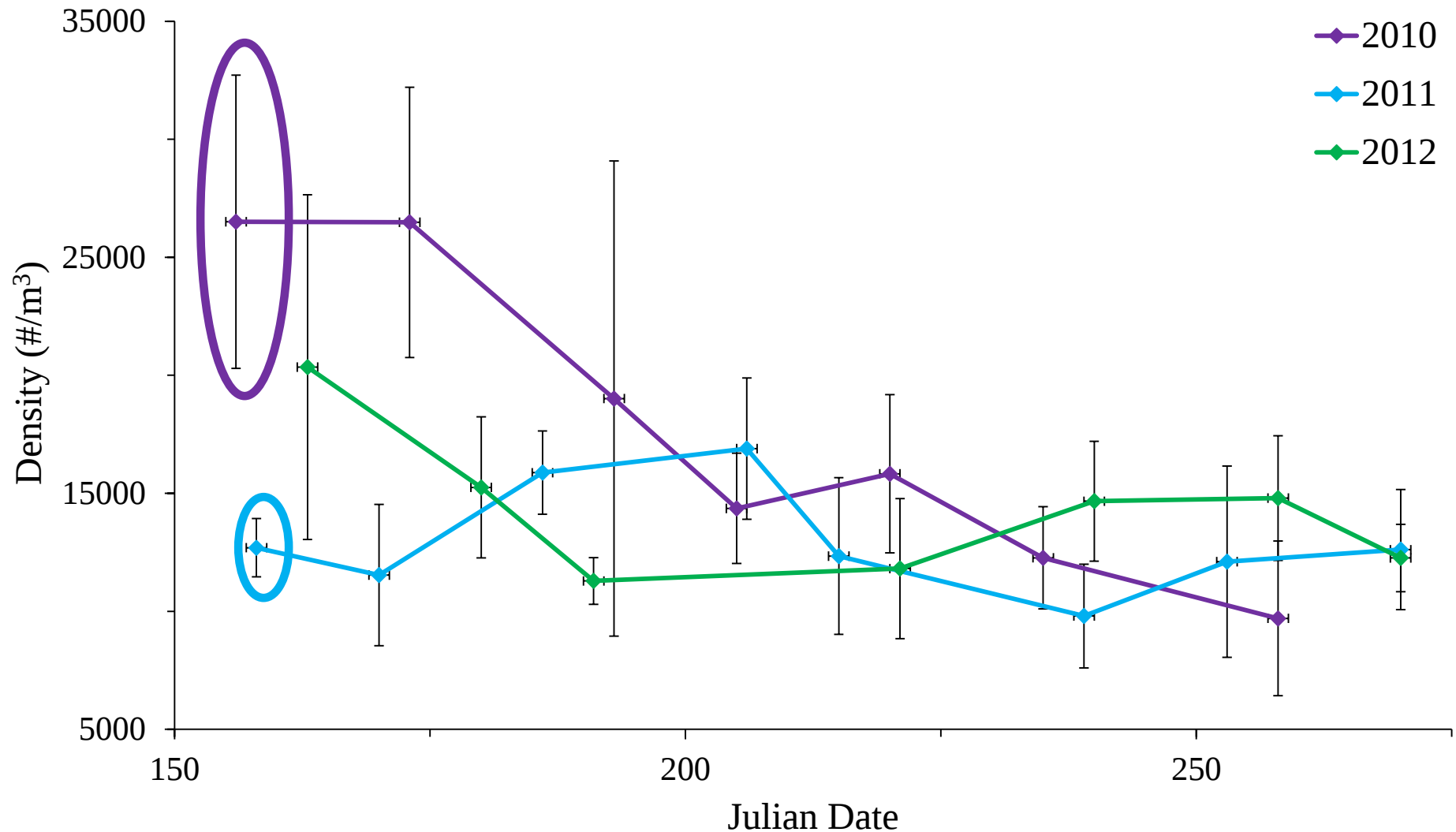
Results- Secchi depth



Results- chlorophyll-*a* concentrations



Results- total zooplankton density

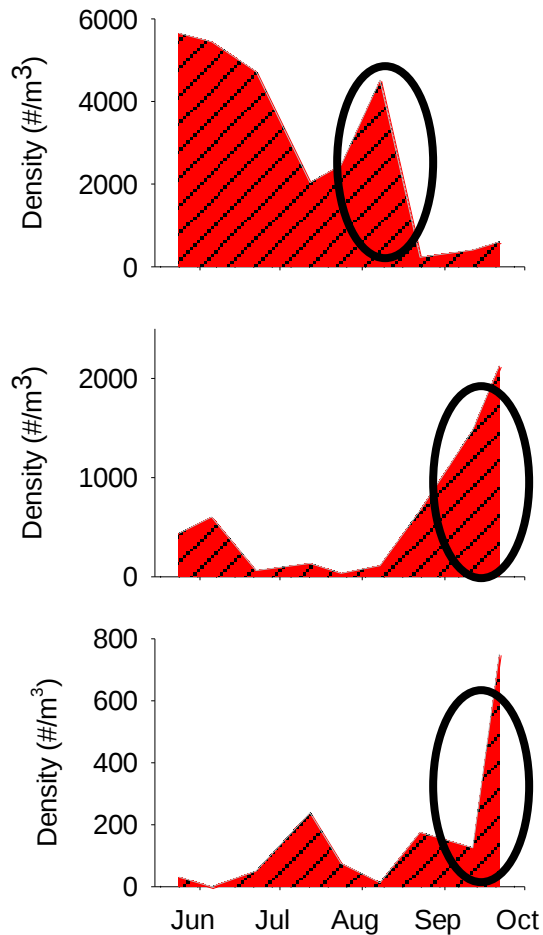


Types of responses

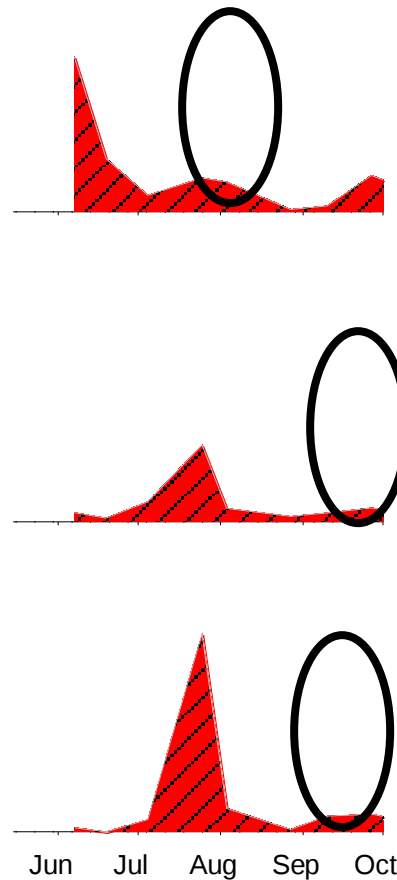
Immediate and lasting negative response

Category	Taxon	Response	2010	2011	2012
Asplanchnid rotifer	<i>Asplanchna priodonta</i>	Immediate and lasting disadvantage	^	v	v
Brachionid rotifer	<i>Keratella cochlearis</i>		^	v	v
Synchaetid rotifer	<i>Polyarthra spp.</i>		^	v	v
Daphnid cladoceran	<i>Daphnia retrocurva</i>	Delayed disadvantage	^	^	v
Calanoid copepod	Diaptomidae		^	^	v
Cyclopoid copepod	<i>Mesocyclops edax</i>		>	^	v
Conochilid rotifer	<i>Conochilus unicornis</i>	Immediate disadvantage	^	v	^
Synchaetid rotifer	<i>Ploesoma spp.</i>		^	v	^
Daphnid cladoceran	<i>Ceriodaphnia reticulata</i>	Delayed advantage	v	v	^
Bosminid cladoceran	<i>Eubosmina coregoni</i>		v	v	^
Cyclopoid copepod	<i>Tropocyclops prasinus mexicanus</i>		>	v	^

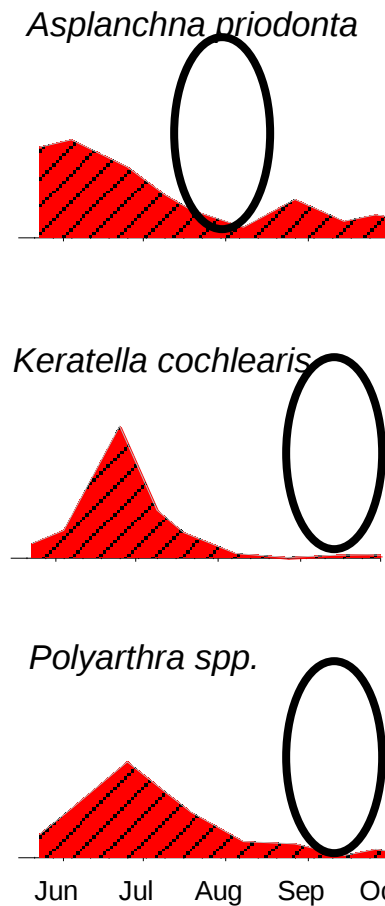
Immediate and lasting negative response



2010



2011



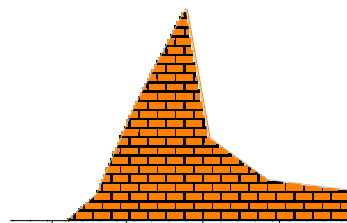
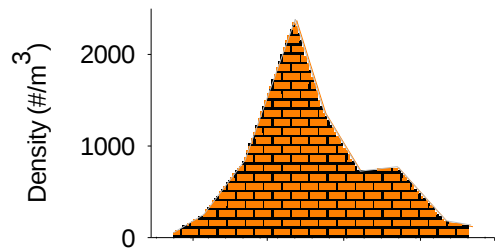
2012



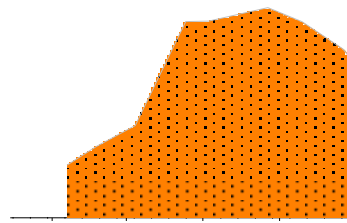
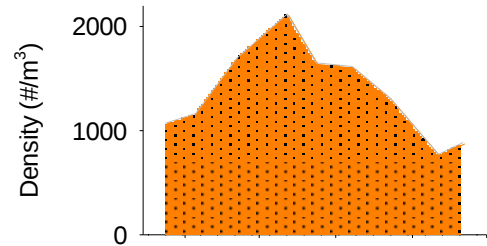
Delayed negative response

Category	Taxon	Response	2010	2011	2012
Asplanchnid rotifer	<i>Asplanchna priodonta</i>	Immediate and lasting disadvantage	^	v	v
Brachionid rotifer	<i>Keratella cochlearis</i>		^	v	v
Synchaetid rotifer	<i>Polyarthra spp.</i>		^	v	v
Daphnid cladoceran	<i>Daphnia retrocurva</i>	Delayed disadvantage	^	^	v
Calanoid copepod	Diaptomidae		^	^	v
Cyclopoid copepod	<i>Mesocyclops edax</i>		>	^	v
Conochilid rotifer	<i>Conochilus unicornis</i>	Immediate disadvantage	^	v	^
Synchaetid rotifer	<i>Ploesoma spp.</i>		^	v	^
Daphnid cladoceran	<i>Ceriodaphnia reticulata</i>	Delayed advantage	v	v	^
Bosminid cladoceran	<i>Eubosmina coregoni</i>		v	v	^
Cyclopoid copepod	<i>Tropocyclops prasinus mexicanus</i>		>	v	^

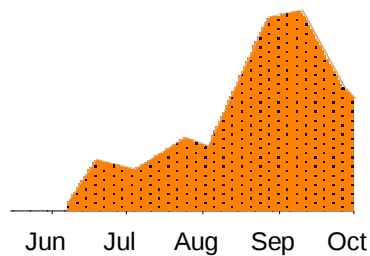
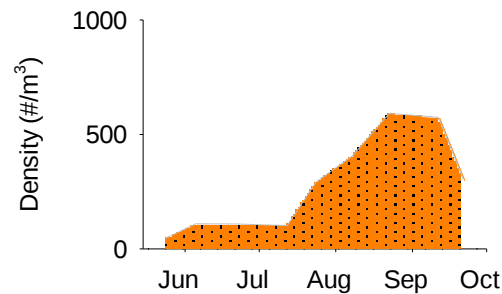
Delayed negative response



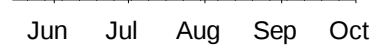
Daphnia retrocurva



Diaptomidae



Mesocyclops edax



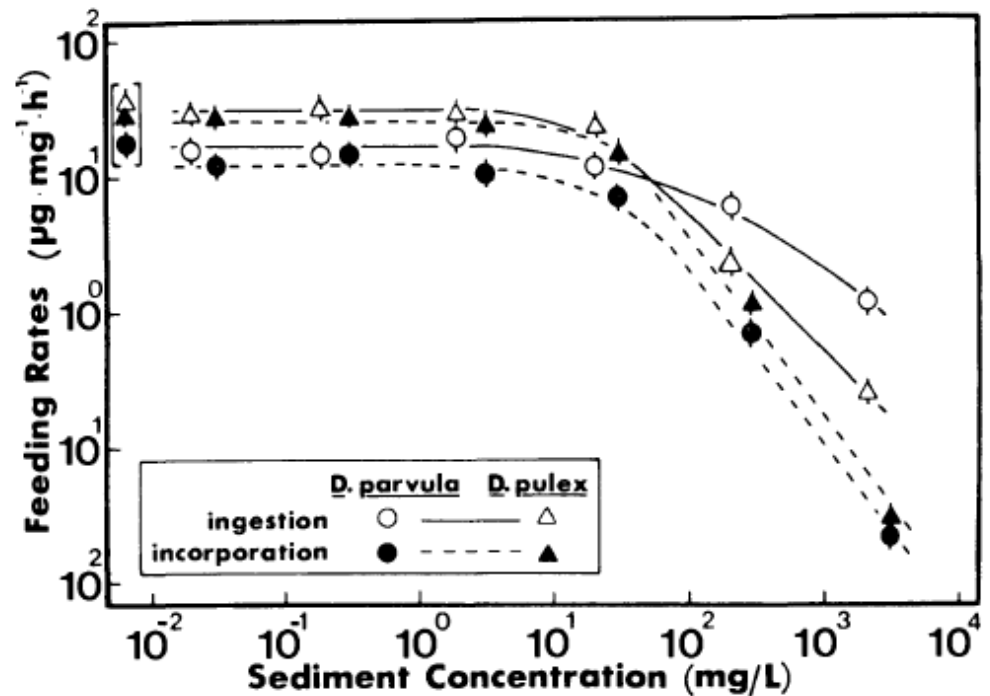
2012



Daphnia and turbidity



- *Daphnia* feeding rates decrease as sediment concentration increases

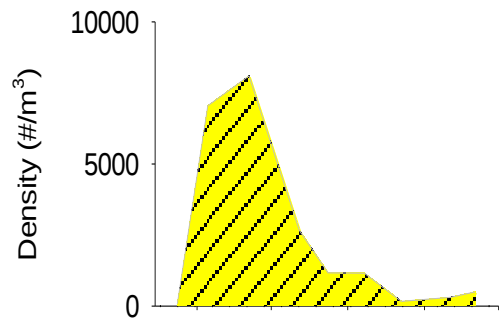


Arruda et al. 1983

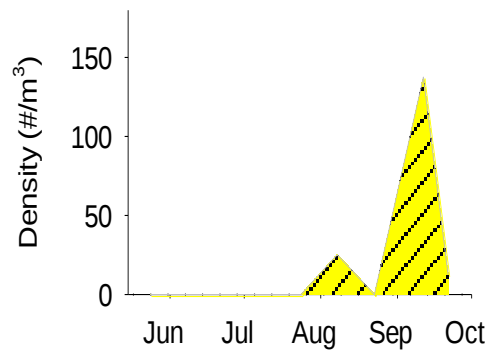
Immediate negative response

Category	Taxon	Response	2010	2011	2012
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Brachionid rotifer	<i>Keratella cochlearis</i>		^	v	v
Synchaetid rotifer	<i>Polyarthra spp.</i>		^	v	v
Daphnid cladoceran	<i>Daphnia retrocurva</i>	Delayed disadvantage	^	^	v
Calanoid copepod	Diaptomidae		^	^	v
Cyclopoid copepod	<i>Mesocyclops edax</i>		>	^	v
Conochilid rotifer	<i>Conochilus unicornis</i>	Immediate disadvantage	^	v	^
Synchaetid rotifer	<i>Ploesoma spp.</i>		^	v	^
Daphnid cladoceran	<i>Ceriodaphnia reticulata</i>	Delayed advantage	v	v	^
Bosminid cladoceran	<i>Eubosmina coregoni</i>		v	v	^
Cyclopoid copepod	<i>Tropocyclops prasinus mexicanus</i>		>	v	^

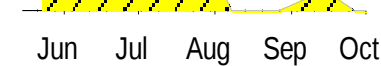
Immediate negative response



Conochilus unicornis



Ploesoma spp.



2010

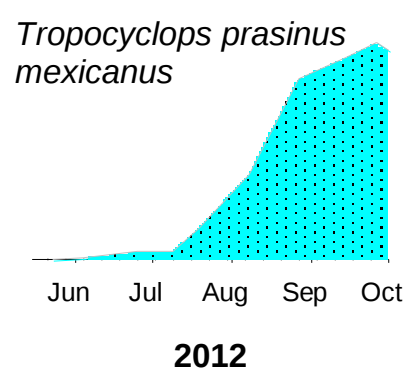
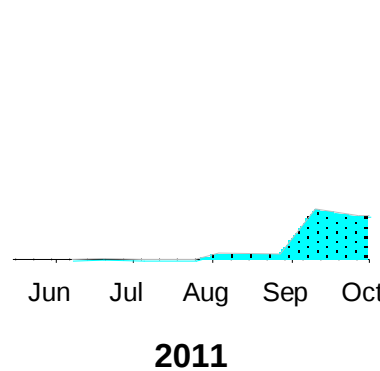
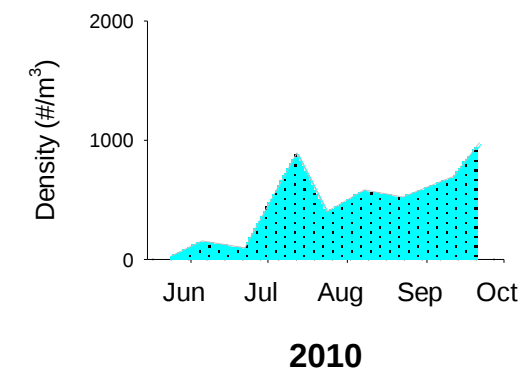
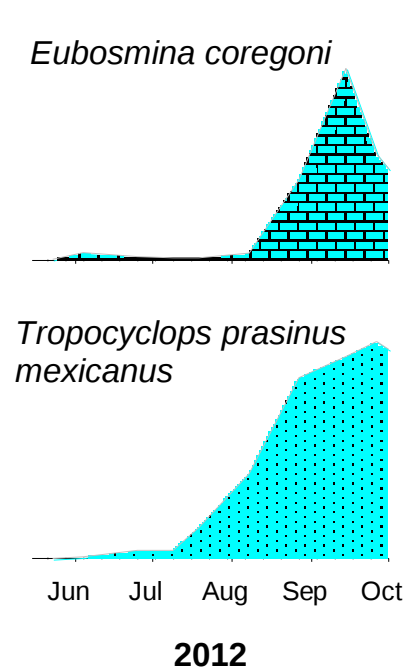
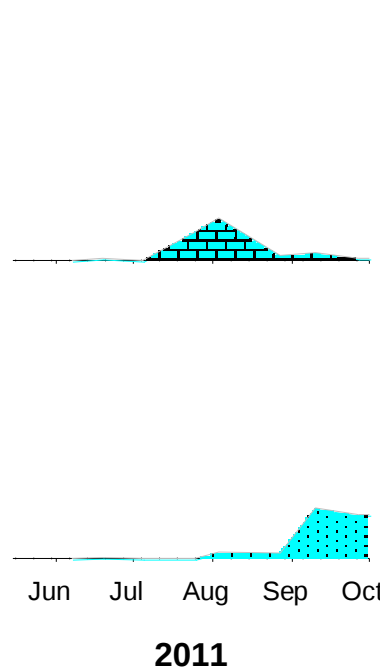
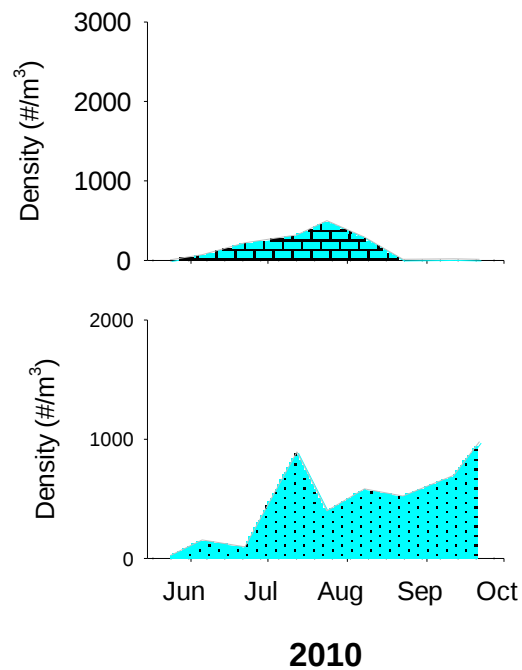
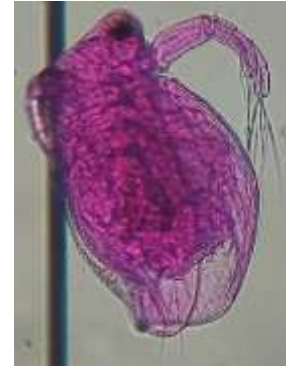
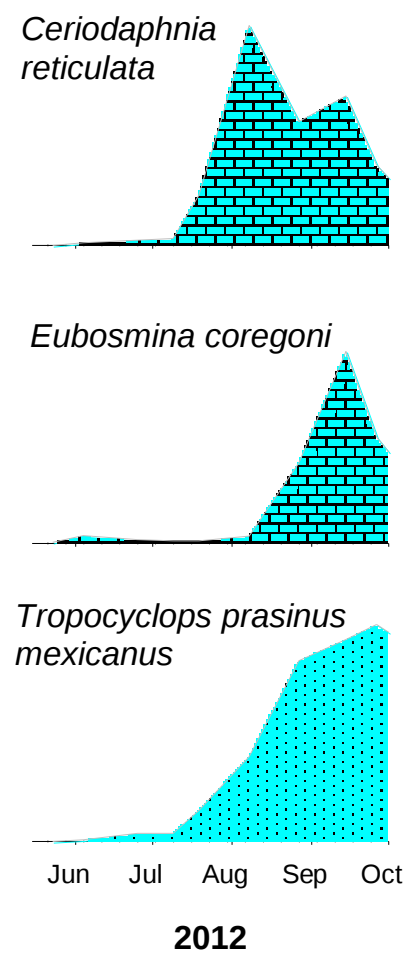
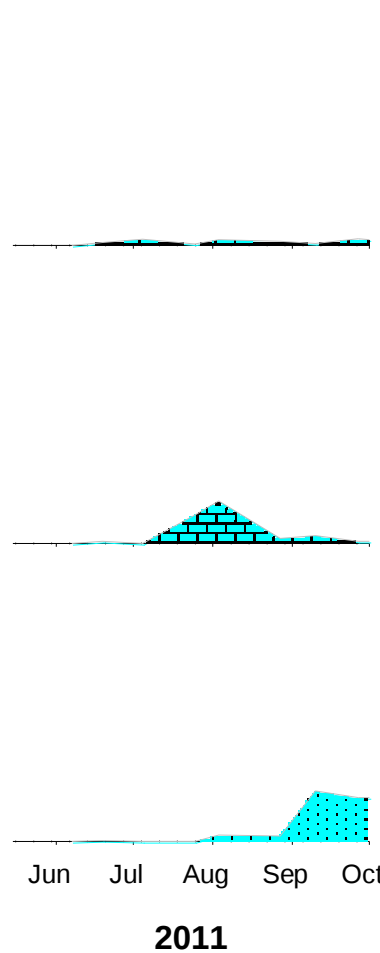
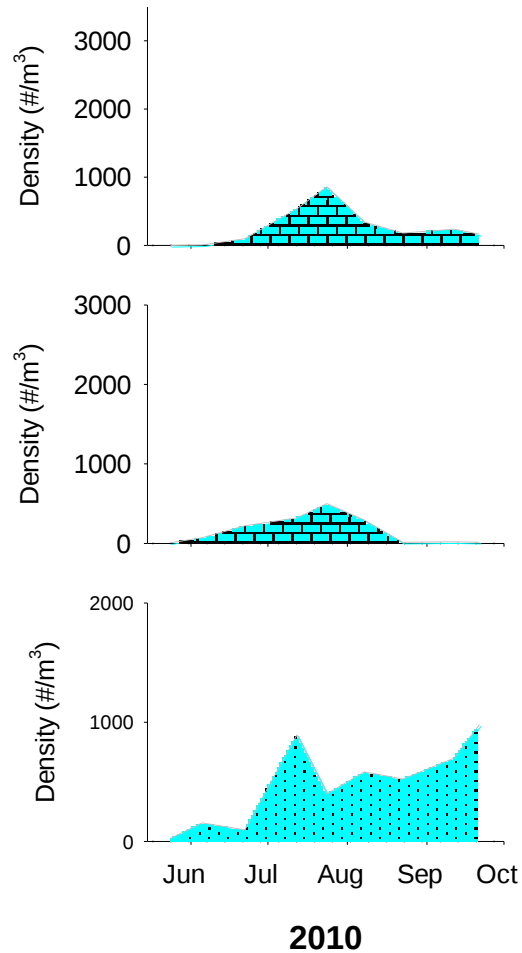
2011

2012

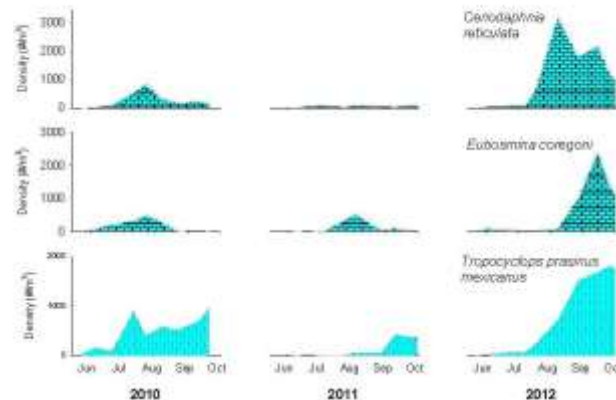
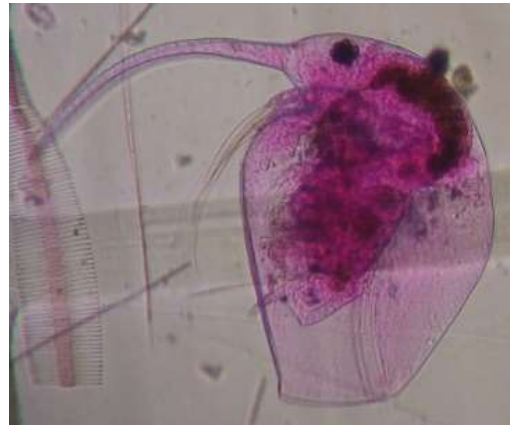
Delayed positive response

Category	Taxon	Response	2010	2011	2012
Asplanchnid rotifer	<i>Asplanchna priodonta</i>	Immediate and lasting disadvantage	^	v	v
Brachionid rotifer	<i>Keratella cochlearis</i>		^	v	v
Synchaetid rotifer	<i>Polyarthra spp.</i>		^	v	v
Daphnid cladoceran	<i>Daphnia retrocurva</i>	Delayed disadvantage	^	^	v
Calanoid copepod	Diaptomidae		^	^	v
Cyclopoid copepod	<i>Mesocyclops edax</i>		>	^	v
Conochilid rotifer	<i>Conochilus unicornis</i>	Immediate disadvantage	^	v	^
Synchaetid rotifer	<i>Ploesoma spp.</i>		^	v	^
Daphnid cladoceran	<i>Ceriodaphnia reticulata</i>	Delayed advantage	v	v	^
Bosminid cladoceran	<i>Eubosmina coregoni</i>		v	v	^
Cyclopoid copepod	<i>Tropocyclops prasinus mexicanus</i>		>	v	^

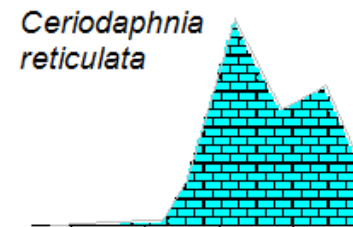
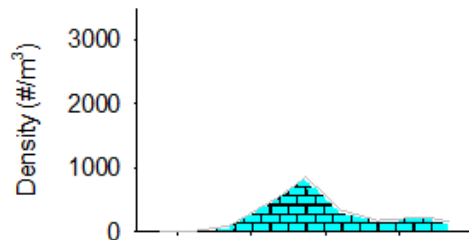
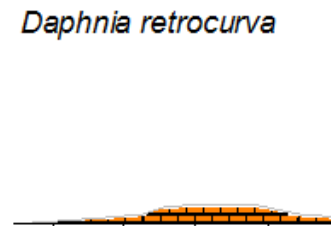
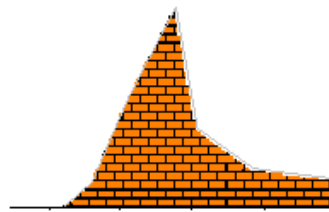
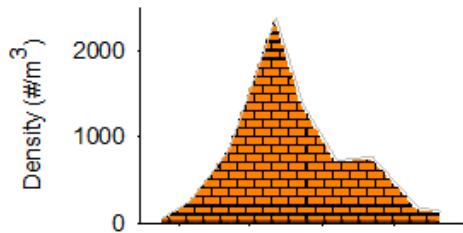
Delayed positive response



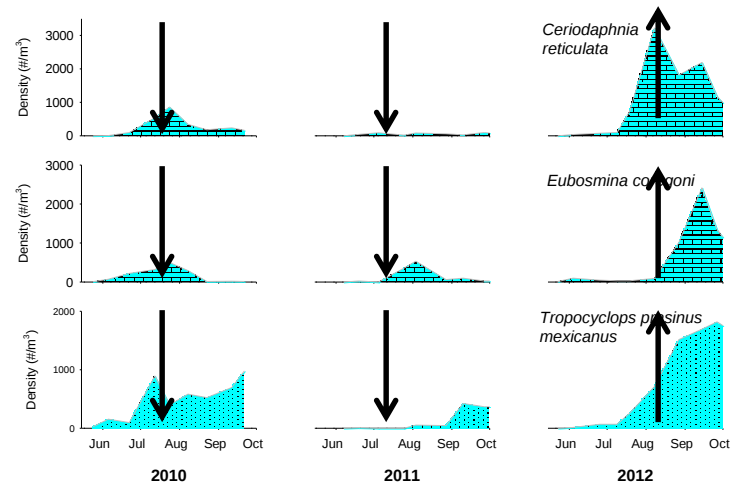
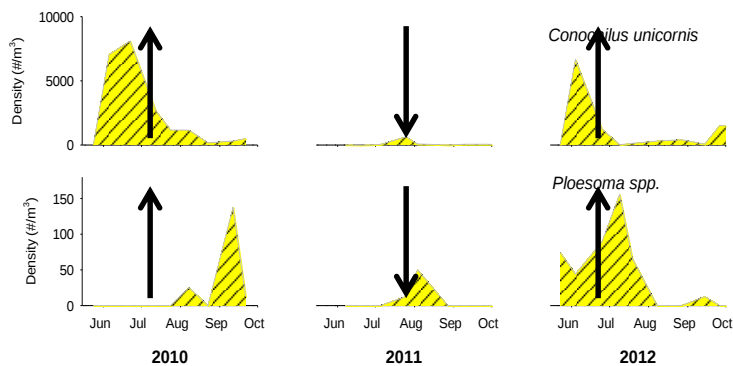
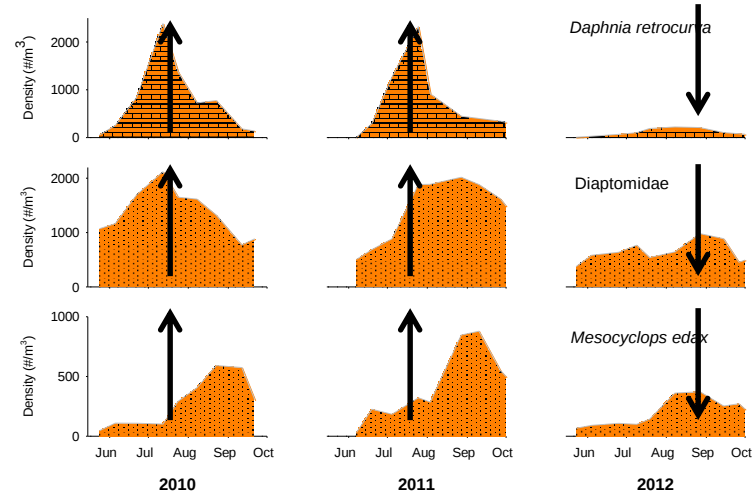
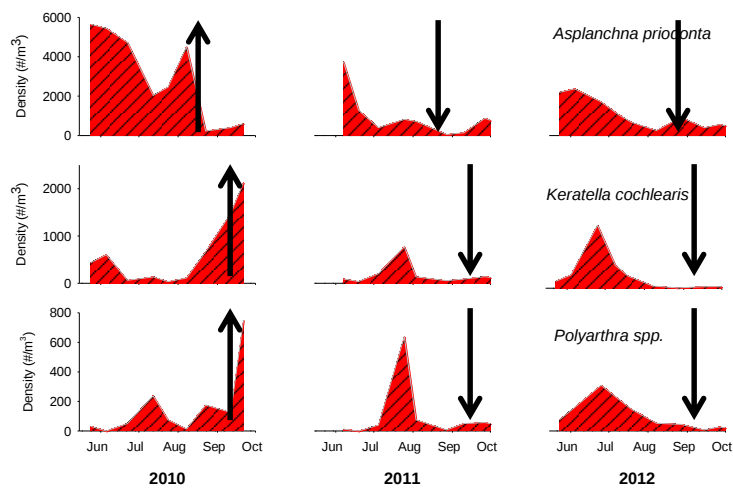
Who we might see more of:



Ceriodaphnia vs. *Daphnia*



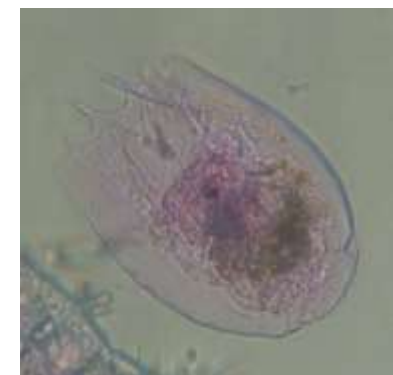
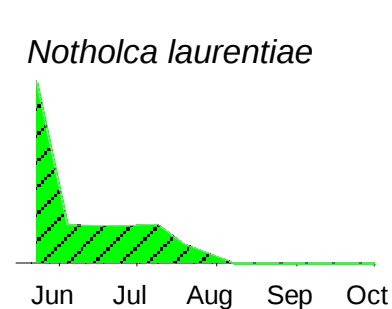
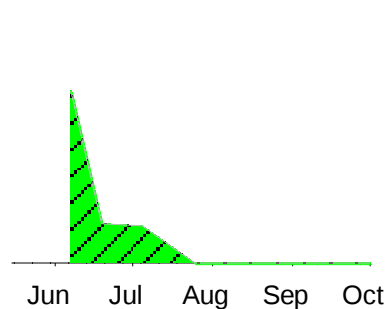
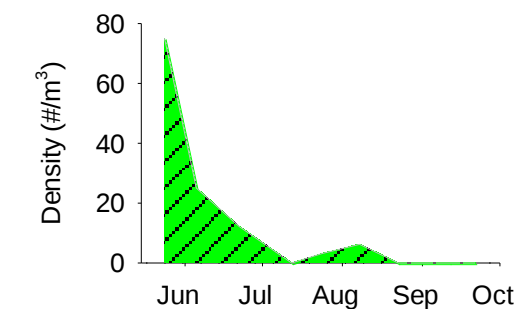
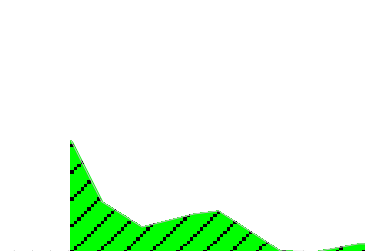
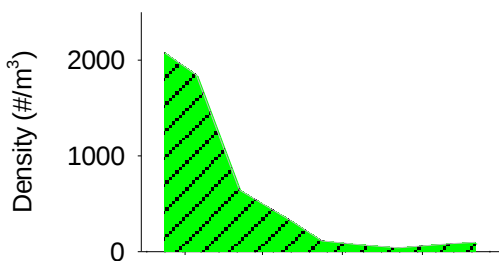
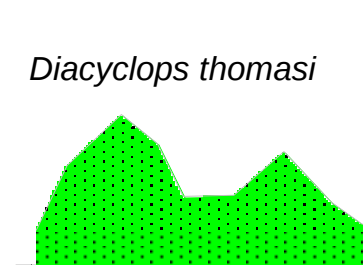
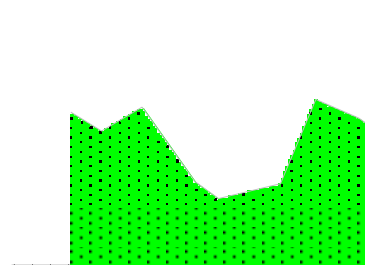
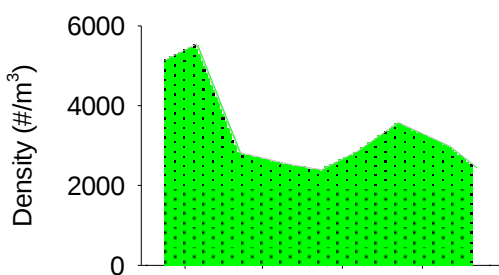
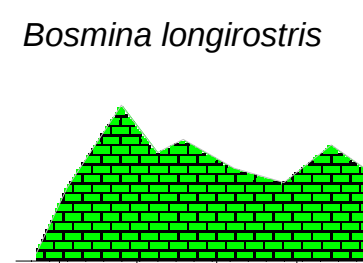
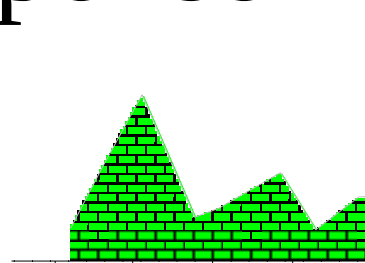
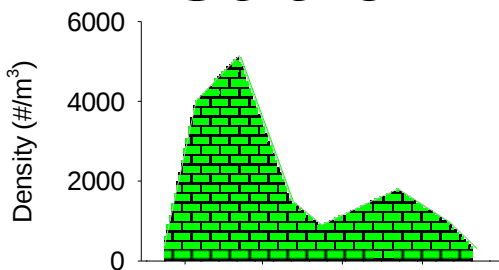
So, everything's always changing, right?



Neutral response

Category	Taxon	Response	2010	2011	2012
Asplanchnid rotifer	<i>Asplanchna priodonta</i>	Immediate and lasting disadvantage	^	v	v
Brachionid rotifer	<i>Keratella cochlearis</i>		^	v	v
Synchaetid rotifer	<i>Polyarthra spp.</i>		^	v	v
Daphnid cladoceran	<i>Daphnia retrocurva</i>	Delayed disadvantage	^	^	v
Calanoid copepod	Diaptomidae		^	^	v
Cyclopoid copepod	<i>Mesocyclops edax</i>		>	^	v
Conochilid rotifer	<i>Conochilus unicornis</i>	Immediate disadvantage	^	v	^
Synchaetid rotifer	<i>Ploesoma spp.</i>		^	v	^
Bosminid cladoceran	<i>Bosmina longirostris</i>	Neutral response	>	>	>
Cyclopoid copepod	<i>Diacyclops thomasi</i>		>	>	>
Brachionid rotifer	<i>Kellicottia longispina</i>		>	>	>
Brachionid rotifer	<i>Notholca laurentiae</i>		>	>	>
Daphnid cladoceran	<i>Ceriodaphnia reticulata</i>	Delayed advantage	v	v	^
Bosminid cladoceran	<i>Eubosmina coregoni</i>		v	v	^
Cyclopoid copepod	<i>Tropocyclops prasinus mexicanus</i>		>	v	^

Neutral response



2010

2011

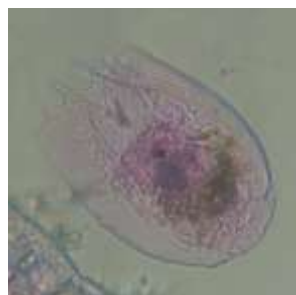
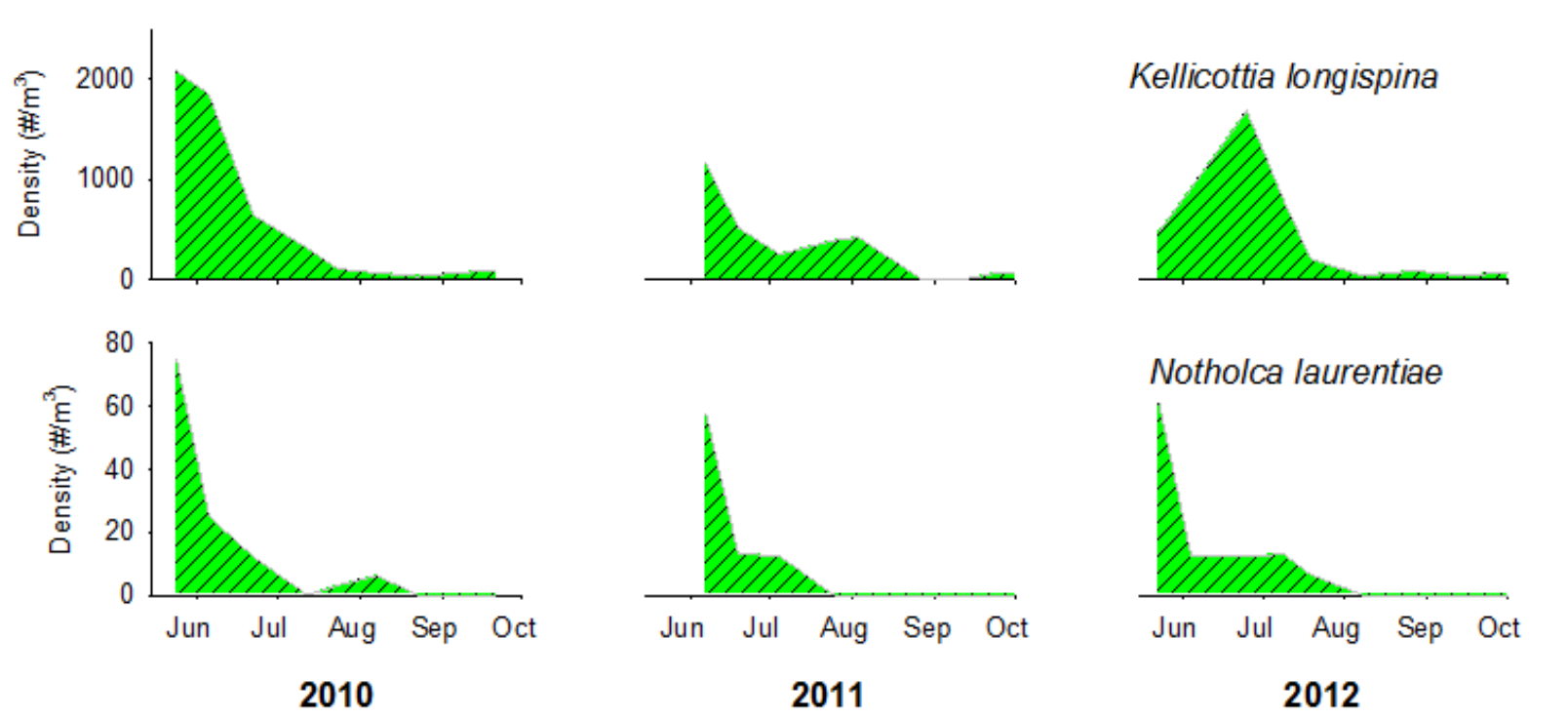
2012



Diacyclops thomasi

the dominant copepod since 1930

Is early seasonality an advantage in flood years?



Summary

– *Long-term patterns:*

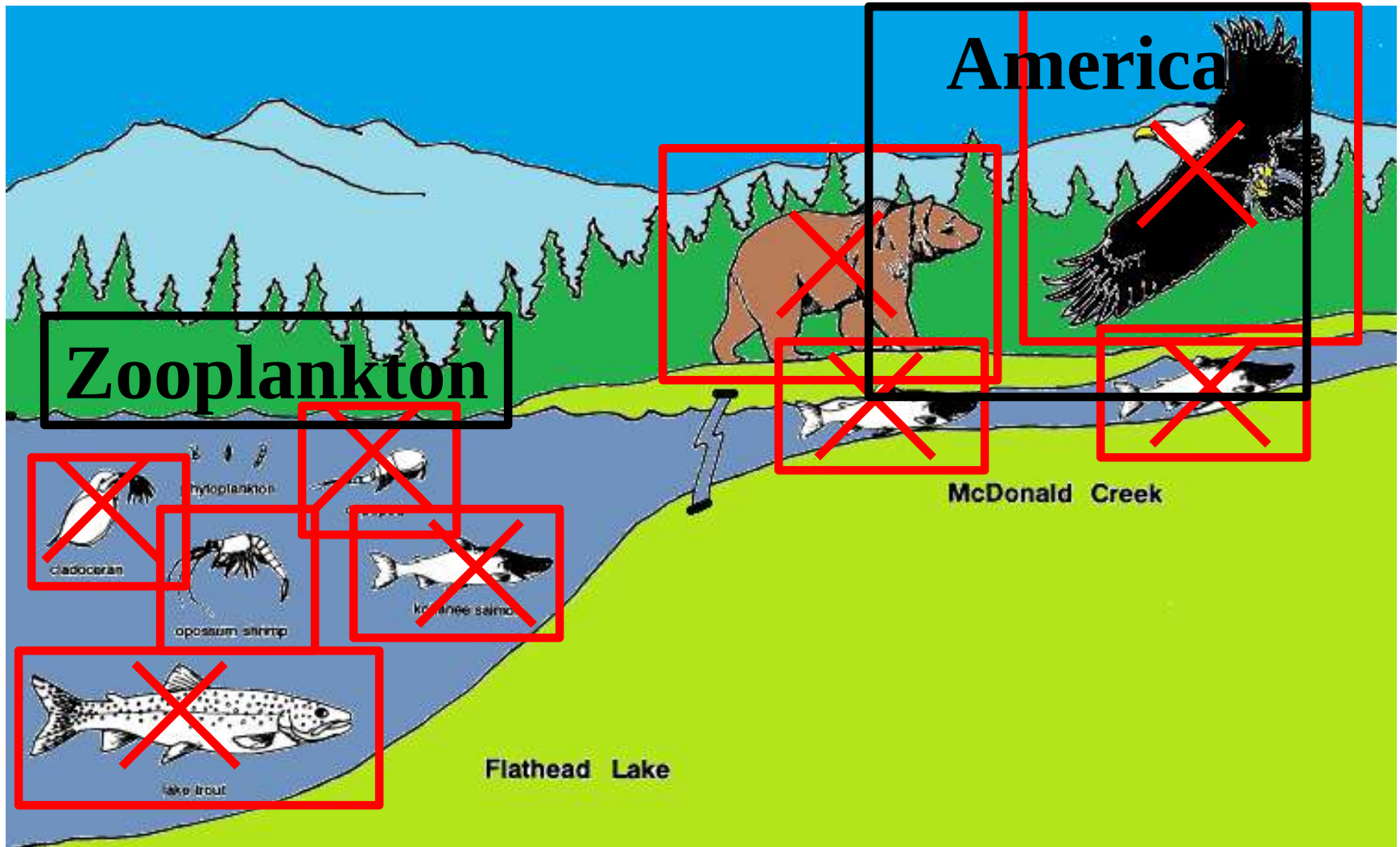
Rotifer declines in richness and abundance- Mid 1990s

- Mysid declines → Zebra Mussel, Rotifer relationships
- Phytoplankton community shifts
 - » long-term climate change?

--- *Flooding / climate change:*

Community shifts based on autecological responses.

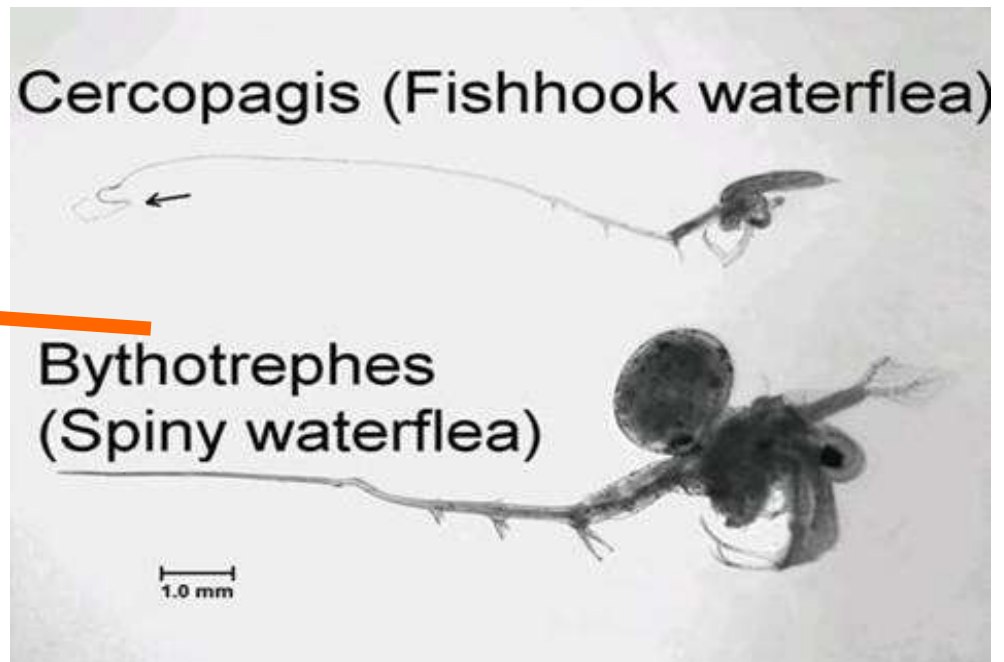
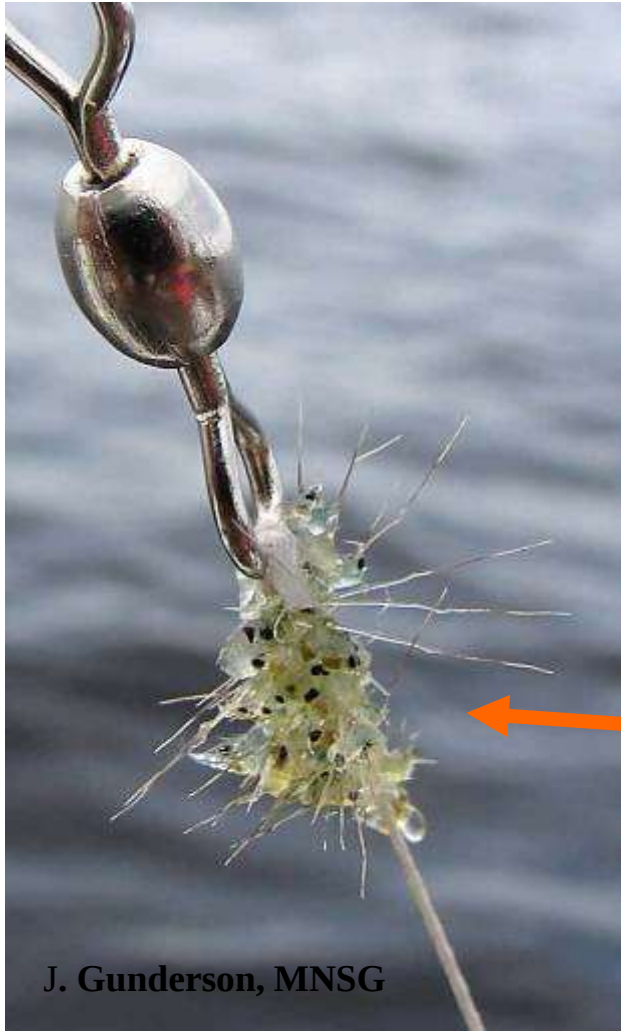
Why you should care about plankton



Questions?



Future Invasive Zooplankton





Lake George= Spiny waterflea source





Lake Champlain Food Web



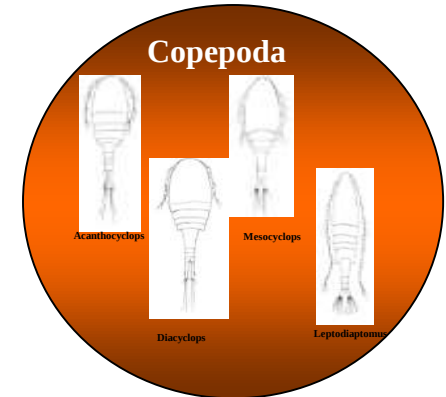
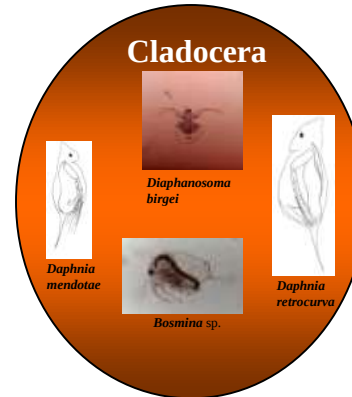
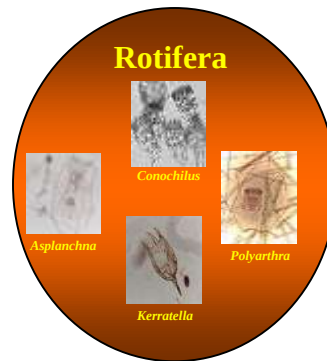
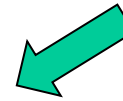
Mysis diluviana



Leptodora



Bythotrephes





STOP AQUATIC HITCHHIKERS!

Prevent the transport of nuisance species.
Clean all recreational equipment.
www.ProtectYourWaters.net

Aquatic Invaders to Look Out For:



B. Smith, ADPP

Water Chestnut

- Triangular leaves & toothed edges
- Forms dense floating mats
- Seeds & plants attach to trailers



J.E. Manning, UVM & D. Bels

Zebra/Quagga Mussel

- Tiny, "D" or oval shaped striped shells
- Covers hard surfaces & sharp shells cut feet
- Shells attach to plants & boat bottoms



A. Fox, Regenering & Managing UVM

Eurasian Watermilfoil

- Feathery leaves in whorls of 4
- Forms dense beds
- Plant fragments attach to boats & trailers



C.P. Wu & A. Long-Akers, SUNY Oneonta

Spiny Waterflea

- Tiny, 1/2" crustacean with long, barbed tail
- Competes with fish for food
- Masses collect on fishing line



D. Mendenhall, E. Yantis, Regenering UVM

Hydrilla

- Blade-like leaves in whorls of 4-8
- Forms dense beds
- Plant fragments attach to boats & trailers



J. Gendron & D. Bels, Regenering UVM

Live Bait

- Non-native crayfish and minnows
- Alters aquatic habitats & may carry pathogens
- Often released or illegally stocked

The Watershed Stewardship Program (WSP) is a collaborative education program offered at water access sites in the Adirondack and Champlain regions to prevent the spread of aquatic invasive species and help preserve the integrity of water resources and local economies.



Adirondack Park
Invasive Plant Program



Lake George
Association



Lake Champlain
Basin Program

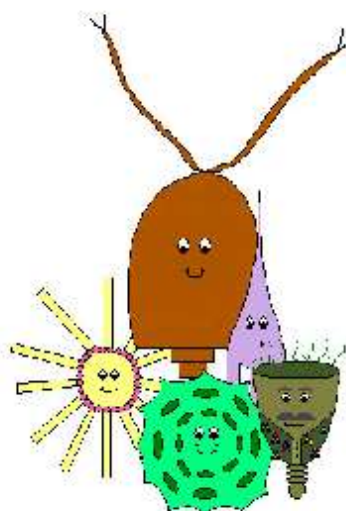


Adirondack Watershed
Institute of Paul Smith's
College



NYS Dept. of
Environmental
Conservation

ADVENTURES OF CAPTAIN FISHHOOK WATERFLEA



THE INVASION OF LAKE CHAMPLAIN

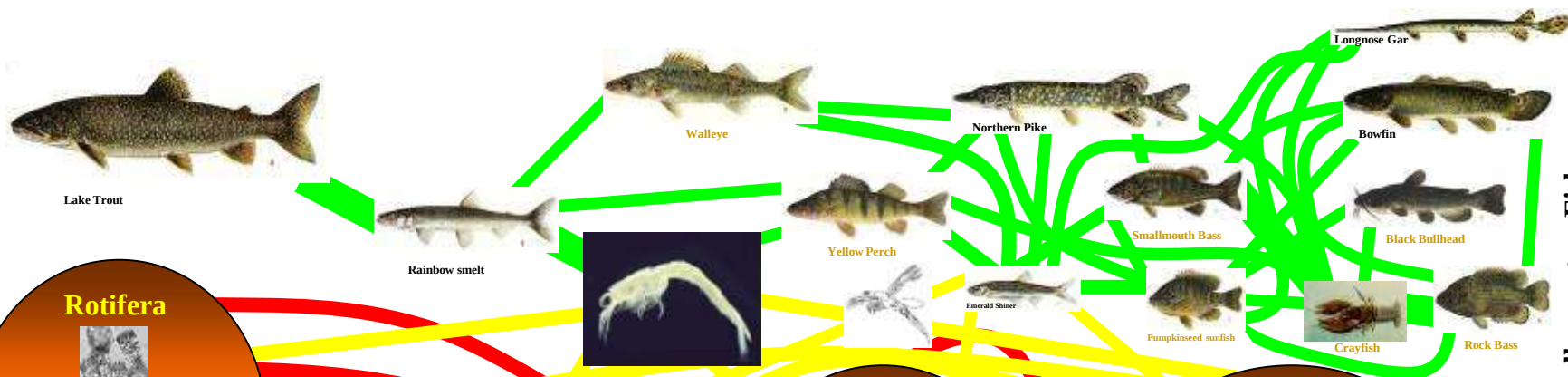
By Alexandra Mihalek, Carrienne Pershyn and Casey
Binggeli (Edited by Dr. Timothy B. Mihuc)



Lake Champlain Food Web



Deep water: Fish



Shallow water: Fish

Rotifera

Asplanchna
Conochilus
Polyarthra
Keratella

Desmids

Closterium
Scenedesmus

Protozoa

Ciliates
Dinoflagellates

Diatoms

Pinnularia
Rhizosolenia
Fragilaria
Navicula

Blue-Green Algae

Anabaena
Microcystis
Cyanothece

Copepoda

Acanthocyclops
Mesocyclops
Diacyclops
Leptodiamydus

Green Algae

Chlorella
Volvox
Plectonicon

BENTHOS
Oligochaeta – Chironomidae – Hexagenia sp.
Bacterial-Detrital Complex

Sturgeon
Channel Catfish
Sculpin

Alewife

- Marine member of herring family
- Discovered in Lake St. Catherine in 1997
- Compete with native fish for food
- Preys directly on eggs and young native fish species
- Interferes with trout & salmon reproduction
- Unstable forage base, periodic die-offs
- Quebec – Aug 2003
- Vermont – July 2004
- Management options?



Lake Champlain Region

